



Which Natural Products Merit Development as Insecticides? Prioritizing Candidates through Mechanism, Selectivity, Synergy, Chemical Stability, and Resistance Risk

Mateo Alvarez^{1*}, Sofia Herrera², Diego Cruz¹, Helena Costa³

¹Department of AI-Based Renewable Energy Systems, Faculty of Engineering, National University of Colombia, Bogota, Colombia.

²Department of Energy Data Intelligence, Faculty of Engineering, University of Chile, Santiago, Chile.

³Department of Artificial Intelligence in Energy Engineering, Faculty of Engineering, University of Porto, Porto, Portugal.

ABSTRACT

Natural products offer substantial chemical diversity for insect-control discovery, but bioactivity screening generates many more promising observations than development-ready candidates. The central difficulty is therefore not merely identifying plant-derived materials that affect insects, but deciding which chemically defined candidates justify continued investment. Acute laboratory potency is often privileged even when chemical identity, target relevance, selectivity, mixture interactions, environmental stability, formulation feasibility, and resistance liability remain uncertain. This article develops an original, explicitly non-validated prioritization structure that integrates these domains into a staged decision logic for natural-product insecticides. The approach distinguishes discovery evidence from development evidence and treats candidate value as conditional on the intended pest, life stage, exposure route, production system, formulation, environmental setting, and non-target compartments. The synthesis indicates that credible candidates combine reproducible chemical definition with biologically relevant activity, mechanism-supported but not overclaimed target evidence, an exposure-aware selectivity profile, manageable stability and delivery characteristics, and a resistance assessment that considers both the active substance and the complete formulated product. Synergy may increase activity or reduce the required amount of one component, but it does not inherently lower resistance risk and may enlarge uncertainty when interaction mechanisms, component ratios, or non-target responses are insufficiently characterized. Major limitations include heterogeneous assay designs, incomplete chemical standardization, limited cross-species testing, and weak linkage between laboratory effects and realistic use conditions. The principal implication is that candidate advancement should occur through sequential evidence gates rather than a potency-led ranking. Such a structure can improve transparency in research investment while remaining subject to prospective validation across chemically, taxonomically, and operationally diverse insect-control systems.

Keywords: Botanical insecticides, Bioactivity-guided discovery, Plant source selection, Chemical identity, Chemotype, Active constituent.

HOW TO CITE THIS ARTICLE: Alvarez M, Herrera S, Cruz D, Costa H. Which Natural Products Merit Development as Insecticides? Prioritizing Candidates through Mechanism, Selectivity, Synergy, Chemical Stability, and Resistance Risk. Entomol Appl Sci Lett. 2025;12(3):1-12. <https://doi.org/10.51847/sgwgGwFPZt>

Corresponding author: Mateo Alvarez

E-mail ✉ malvarez@unal.edu.co

Received: 10/03/2025

Accepted: 29/06/2025

INTRODUCTION

Natural products occupy an important but difficult position in insecticide discovery. They provide diverse molecular scaffolds, multi-

constituent mixtures, volatile compounds, feeding deterrents, growth disruptors, neuroactive substances, and other biologically active materials that may support direct pest control or inspire semisynthetic development.

Nevertheless, the volume of reported botanical activity has not produced a corresponding number of consistently formulated and widely adopted products. Botanical bioactivity becomes developmentally meaningful only when chemical reproducibility, practical efficacy, safety, and delivery can be demonstrated together [1]. The relevant question is therefore not whether a plant extract, essential oil, metabolite, or derivative can affect an insect under at least one experimental condition. It is whether the candidate possesses a sufficiently coherent combination of chemical, biological, technological, and risk-related properties to justify progression.

This distinction separates discovery value from development priority. Natural products may function as direct active ingredients, sources of optimized derivatives, synergists, repellents, or templates for identifying insect-relevant targets. Each route, however, carries different requirements for chemical supply, analytical control, formulation, exposure, toxicology, intellectual development, and resistance management. Natural products provide both direct pesticidal actives and chemical starting points, yet their development value depends on properties extending well beyond initial potency [2]. A compound with moderate activity but reproducible chemistry, a plausible delivery route, and a credible safety margin may be more developable than a highly potent extract whose active constituents, source variability, persistence, or non-target effects remain unresolved.

The natural origin of a candidate does not resolve these uncertainties. Botanical materials may vary with genotype, chemotype, plant organ, developmental stage, geography, cultivation, storage, extraction procedure, and analytical definition. Their constituents can volatilize, oxidize, photodegrade, interact antagonistically or synergistically, or change biological availability after formulation. The same intervention may consequently produce different target and non-target effects across laboratory, greenhouse, storage, aquatic, soil, and field contexts. A sustainable botanical insecticide cannot be inferred from plant origin alone because composition, delivery, persistence, and environmental exposure jointly shape its performance and risk [3]. Sustainability is

therefore an evidence-dependent property of a defined product and use pattern, not an intrinsic property conferred by botanical provenance.

This article addresses the absence of an integrated decision structure linking early natural-product discovery to insecticide development priority. Its purpose is not to present a validated scoring system, prescribe universal thresholds, or claim that every candidate should follow an identical commercial pathway. Instead, it organizes the selected evidence around seven connected questions: whether the candidate is chemically and biologically reproducible; whether its mechanism is sufficiently characterized and relevant to the intended target; whether its selectivity is credible under plausible exposure; whether synergy, stability, and formulation improve or complicate its profile; whether resistance and cross-resistance liabilities are understood; which criteria should govern progression; and which studies are needed to reduce the most decision-relevant uncertainties. The central argument is that development priority should be assigned through staged evidence integration. In vitro potency is not equivalent to development priority, mechanistic novelty is not equivalent to target validation, natural origin is not equivalent to selectivity, and synergy in a mixture is not equivalent to reduced resistance risk.

The candidate-selection problem in natural-product discovery

Natural-product discovery commonly begins with ethnobotanical knowledge, whole-plant materials, crude extracts, essential oils, fractions, isolated metabolites, or structurally related derivatives. These entry points are not interchangeable. Traditional use can identify biologically plausible plants and locally relevant application practices, but it may involve variable material, informal preparation, uncertain dose, multiple modes of exposure, or context-specific effectiveness. Traditional or local pesticidal use is a valuable discovery signal, but it does not by itself establish a reproducible, scalable, or safe product candidate [4]. Candidate selection must therefore preserve the informational value of local use while requiring botanical authentication, defined plant material, traceable processing, chemical profiling, supply feasibility,

and testing under an explicitly stated use context. Otherwise, apparent efficacy may not be reproducible across harvests, production sites, extraction procedures, or intended users.

Potency-centred screening creates a second problem. A low lethal concentration or rapid laboratory response may appear to provide a convenient ranking variable, yet candidates tested under different routes, durations, life stages, carriers, environmental conditions, or endpoints cannot be ordered credibly through potency alone. An impressive laboratory response remains a screening result until it survives realistic exposure, formulation, persistence, and field-performance tests [5]. Development decisions must also account for effects that acute mortality assays do not capture. The biorational label does not remove the need to examine sublethal effects, behavioral disruption, life-stage sensitivity, and ecological consequences [6]. Reduced feeding, impaired development, repellency, delayed reproduction, altered movement, or disruption of beneficial-organism performance may be relevant outcomes, but their interpretation depends on whether they improve pest suppression, merely redistribute insects, or create unintended effects elsewhere in the system.

A defensible candidate-selection process should therefore begin with an intended product profile rather than with a decontextualized list of active plants. The target insect, life stage, crop or stored commodity, application route, exposure duration, required persistence, user setting, and principal non-target compartments should be specified before comparing candidates. Candidate ranking should be anchored to a defined pest, life stage, exposure route, and intended use rather than to generic claims of biopesticidal activity [7]. Within that context, chemical identity, mechanism, selectivity, formulation, stability, and resistance relevance become connected decision dimensions rather than independent desirable attributes. Recent syntheses converge that commercial progress depends on standardization, realistic efficacy, safety, delivery, and manufacturability rather than assay potency alone [1, 2]. This convergence supports a multidimensional selection logic, but it does not establish universal weights or prove that a single candidate profile will be optimal across production systems.

The evidence dimensions and interpretive boundaries for candidate-selection problem in natural-product discovery are summarized in **Table 1**.

Table 1. The Candidate-Selection Problem in Natural-Product Discovery: Chemical Identity, Mechanism, Standardization, Formulation, Safety, Resistance, and Translation Requirements

Candidate or product factor	Chemical or biological basis	Required characterization	Potential contribution	Formulation implication	Safety implication	Resistance implication	Development boundary
Botanical source and plant material	Genotype, chemotype, organ, growth stage, environment, and processing can alter composition	Taxonomic authentication, plant part, provenance, harvest conditions, storage, and processing history	Establishes traceability and enables reproducible sourcing	Source variability may require blending, specification limits, or controlled cultivation	Different source profiles may alter hazard and exposure	Variable composition may create inconsistent selection pressure	Traditional use or species identity alone does not define a reproducible candidate
Crude extract or essential oil	Activity may arise from one constituent, several constituents, or nonspecific physicochemical effects	Extraction method, yield, chromatographic profile, major and minor constituents, and batch consistency	Preserves potentially useful multi-constituent activity	Volatility, solubility, oxidation, viscosity, and carrier compatibility affect delivery	Whole-mixture testing is needed because constituent safety cannot simply be added	Multiple constituents do not automatically prevent resistance	Crude-mixture potency does not establish chemical identity, stability, or product consistency
Isolated active constituent	A defined molecule improves attribution of activity and analytical control	Structure confirmation, purity, dose-response, stability, and source or synthesis feasibility	Supports mechanism studies, standardization, and quality control	Isolation may simplify formulation but can remove useful mixture interactions	Defined exposure supports clearer toxicological testing	A single dominant target may create concentrated selection pressure	Isolation does not guarantee superior efficacy, selectivity, or commercial feasibility
Mechanism and target evidence	Effects may involve target engagement, detoxification	Functional assays, biochemical evidence, target	Improves biological plausibility and	Delivery must achieve biologically	Conserved targets or broad modes of action	Mechanistic information can guide	Mechanistic novelty is not

	interference, membrane disruption, behavioral modification, or multiple pathways	engagement, temporal response, and alternative-mechanism testing	informs optimization	relevant exposure at the proposed site of action	may affect beneficial organisms	cross-resistance hypotheses	equivalent to target validation
Target-context efficacy	Biological response depends on species, life stage, route, dose pattern, and environmental setting	Comparative assays using the intended pest, relevant stage, realistic route, and suitable controls	Determines whether activity is relevant to the intended use	Formulation must preserve contact, ingestion, fumigant, or systemic availability as required	Higher effective exposure may also increase non-target contact	Sublethal exposure may intensify selection without delivering adequate control	In vitro or controlled-assay potency is not equivalent to development priority
Standardization and quality control	Batch variation can change efficacy, stability, and risk	Marker compounds, constituent ranges, impurities, degradation products, and validated analytical methods	Enables reproducible manufacture and interpretable testing	Specifications must apply to the formulated product, not only the source extract	Hazard can change when constituent ratios or degradation products vary	Inconsistent batches may create variable selection intensity	A chemically unstandardized material cannot support reliable product-level conclusions
Stability and environmental persistence	Light, oxygen, temperature, moisture, surfaces, and microorganisms may alter active chemistry	Storage stability, photostability, thermal stability, oxidation, release behavior, and residue profile	Identifies whether activity can persist for the required use period	Encapsulation or other delivery systems may protect actives or control release	Increased persistence can extend non-target exposure	Residual low-dose exposure may affect resistance selection	Greater persistence is not automatically preferable
Selectivity and non-target profile	Observed safety reflects intrinsic sensitivity together with route, magnitude, and duration of exposure	Lethal, sublethal, behavioral, reproductive, and functional endpoints across relevant organisms	Supports an exposure-specific benefit–risk judgment	Carriers and delivery systems can alter organismal and environmental exposure	Beneficial arthropods, soil organisms, aquatic biota, and other compartments may require testing	Non-target effects can undermine integrated pest-management value	Natural origin is not equivalent to selectivity
Mixture interaction and synergy	Components may interact through penetration, metabolism, target action, volatility, or delivery	Defined component ratios, appropriate null models, reproducibility, and mechanistic follow-up	May increase activity, broaden effects, or reduce the amount of one component	Formulation can create, amplify, suppress, or destabilize interactions	Mixture safety cannot be inferred solely from component-level tests	Interaction does not inherently diversify resistance selection	Synergy in a mixture is not equivalent to reduced resistance risk
Resistance and cross-resistance liability	Selection depends on target, metabolism, exposure pattern, persistence, and existing resistance mechanisms	Baseline susceptibility, selection studies, inheritance, stability, mechanism, and cross-resistance testing	Clarifies durability and fit with resistance-management strategies	Release profile and residual exposure influence selection conditions	Resistance-management benefits must not be assumed at the expense of safety	Novel origin does not exclude shared detoxification or target-based resistance	Absence of documented resistance is not evidence of low resistance risk
Translation and intended use	Candidate value depends on performance, supply, manufacturing, cost, application, and user constraints	Intended product profile, realistic efficacy, manufacturability, storage, compatibility, and use conditions	Connects discovery evidence to a defined development pathway	Product form must match equipment, handling, storage, and exposure requirements	Use pattern determines which safety evidence is decision-relevant	Operational use determines the actual selection regime	Promising biological activity is not equivalent to a viable insect-control product

Mechanistic novelty and target relevance

Mechanistic evidence can reduce uncertainty about why a natural product affects an insect, but the value of such evidence depends on the chemical definition of the candidate. A crude extract can produce a reproducible phenotype while leaving uncertain whether activity arises

from a dominant constituent, several interacting compounds, degradation products, or general physicochemical stress. Chemical identification coupled to mechanism-focused testing provides stronger candidate evidence than activity attributed only to a crude botanical source [8]. Bioassay-guided fractionation and structural

characterization can therefore strengthen attribution, enable dose-consistent testing, and support comparison across batches. These advantages do not mean that isolated compounds should automatically outrank mixtures. Isolation may remove stabilizing, delivery-enhancing, antagonistic, or synergistic interactions and may produce a candidate that is less practical or less selective than the original material.

The identification of a plausible target should also be separated from validation of its relevance. A receptor, ion channel, enzyme, or developmental pathway may be considered interesting because it is insect-associated, structurally distinctive, or affected under experimental conditions. However, target naming, molecular docking, sequence similarity, or correlation between exposure and pathway disruption does not demonstrate that engagement of that target causes the insecticidal phenotype in the intended pest. Mechanistic novelty should be separated from target validation, which requires functional evidence in the relevant insect rather than target naming or sequence similarity alone [9]. Target relevance becomes stronger when perturbation, pharmacological characterization, biochemical binding, genetic manipulation, rescue experiments, or consistent structure–activity relationships connect target engagement to the observed effect. A natural-product scaffold gains mechanistic credibility when target engagement is supported experimentally, but that evidence remains distinct from proof of operational value [10]. Even a causally supported molecular target does not establish that sufficient exposure will occur in the insect under practical application or that homologous targets in beneficial organisms will remain unaffected.

Mechanistic prioritization should consequently operate as a chain of increasingly demanding claims. The first claim concerns chemical identity: the tested candidate must be sufficiently defined for the effect to be attributed reproducibly. The second concerns mechanism plausibility: the observed temporal, physiological, or biochemical response should be compatible with a proposed pathway. The third concerns causal target relevance in the intended pest. The fourth concerns product-level relevance, including delivery to the site of action, persistence for the required period, selectivity,

and efficacy under realistic conditions. Environmental and formulation variables may alter this chain substantially. When carrier behavior and post-application temperature alter efficacy, the observed outcome cannot be attributed to active-ingredient potency or mechanistic novelty alone [11]. A candidate may therefore possess a biologically interesting mechanism while remaining unsuitable for development because exposure is unstable, formulation dependence is excessive, the target is not sufficiently validated, or the same mechanism creates unacceptable non-target concern.

Selectivity and non-target considerations

Selectivity should be treated as an exposure-specific relationship rather than as an inherent label attached to a natural substance. A candidate may be more toxic to a pest than to one tested beneficial species yet still affect other predators, parasitoids, pollinators, soil organisms, aquatic communities, vertebrates, or ecological processes. Differences in route, timing, body size, metabolism, behavior, target conservation, and internal dose can all shape the apparent selectivity margin. A candidate should not be called selective unless target efficacy is compared with lethal and sublethal responses in relevant beneficial organisms [12]. Such comparisons should use endpoints, exposure periods, and routes that are sufficiently aligned to support interpretation. A large effect difference generated by unrealistic exposure in the pest or negligible exposure in the beneficial organism does not establish intrinsic biological selectivity. Acute mortality is especially inadequate when a non-target organism contributes through behavior, reproduction, predation, parasitism, or population persistence. Compatibility screening should include sublethal outcomes because survival can coexist with altered behavior or performance in a beneficial predator [13]. A surviving natural enemy may consume fewer pests, change movement patterns, reproduce less successfully, or become more vulnerable to subsequent stress. Conversely, a laboratory-detected sublethal effect does not automatically establish loss of biological-control function in the field. Its importance depends on exposure frequency, recovery, behavioral avoidance, population replacement, resource availability,

and the ecological function represented by the measured endpoint. The appropriate interpretation is therefore conditional: sublethal evidence identifies a plausible risk that requires ecological contextualization, whereas absence of acute death cannot be interpreted as evidence of compatibility.

Formulation can further change the selectivity profile by modifying release, persistence, deposition, uptake, and environmental transport. The persistence gained through nano-enabled delivery can extend non-target exposure, making formulation durability a benefit-risk variable rather than an automatically favorable trait [14]. A formulation that protects a volatile active substance may improve target control while also increasing the duration or spatial reach of exposure in aquatic or terrestrial compartments. Selectivity assessment must therefore follow the formulated candidate and intended use pattern rather than stop at the unformulated active ingredient. A defensible selectivity profile must extend beyond the target pest to organisms occupying other exposure compartments, including soil biota [15]. This does not require identical testing of every taxon during early discovery, but it does require a reasoned selection of representative organisms based on application method, environmental fate, ecological function, and plausible exposure. Natural origin is not equivalent to selectivity, and uncertainty about untested compartments is not evidence of no effect.

Synergy, stability, and formulation potential

Natural-product candidates are frequently constrained by low water solubility, volatility, oxidation, photodegradation, thermal sensitivity, limited surface retention, or poor penetration into the target insect. These properties can cause a compound that performs well in a sealed laboratory assay to lose activity rapidly under storage, greenhouse, field, or open-environment conditions. Formulation is therefore not merely a late-stage packaging exercise; it can determine whether the active chemistry reaches the target, remains available for the required interval, and can be applied safely and consistently. Nanoemulsification and related delivery approaches can improve dispersion and stabilize essential-oil preparations, but their developmental value must be judged through

product-level characterization rather than formulation novelty alone [16]. Required evidence includes particle or droplet characteristics where relevant, active-content retention, release behavior, storage stability, compatibility with application equipment, and biological performance after realistic handling. A formulation that increases nominal stability but causes unacceptable persistence, carrier toxicity, manufacturing complexity, or batch variability should not automatically advance.

Synergy presents a related prioritization challenge because multi-constituent natural products are often assumed to derive durability or breadth from chemical complexity. Demonstrated synergy requires more than observing that a mixture is active. The components must be chemically defined, tested individually and in combination, evaluated at explicit ratios, and compared using an appropriate model of expected additivity. Mechanistic work on basil and mandarin essential oils illustrates how mixture activity may reflect coordinated effects on penetration, detoxification, or physiological targets rather than a generic benefit of combining botanical constituents [17]. Such evidence can justify preserving a mixture or designing a formulated combination, but it remains system-specific. Interaction strength may change with component ratio, pest species, life stage, route, temperature, carrier, and endpoint. Antagonism may also occur when one component reduces uptake, accelerates detoxification, changes volatility, or competes at a biological target.

Stability and synergy must consequently be assessed together at the level of the proposed product. Essential-oil nano-biopesticides can retain or modify activity through altered delivery, but the biological outcome remains contingent on formulation composition and exposure context [18]. A mixture that is synergistic immediately after preparation may become additive, antagonistic, or chemically different during storage or after environmental release. Conversely, controlled release may alter the temporal ratio of components even when the initial formulation is chemically correct. The broader synergy literature therefore supports caution: interaction can improve efficacy, reduce the amount of one constituent, or broaden biological effects, but it can also introduce

mechanistic and toxicological uncertainty [19]. Synergy in a mixture is not equivalent to reduced resistance risk. It should be treated as a testable product property whose value depends on reproducibility, stability, exposure, selectivity, and the resistance mechanisms affected by each component.

Resistance liability and cross-resistance

Natural origin does not protect an insecticide candidate from resistance evolution. Selection occurs when heritable variation allows some insects to survive or reproduce under exposure, regardless of whether the active substance is botanical, microbial, semisynthetic, or fully synthetic. Resistance liability depends on the molecular target, detoxification pathways, penetration barriers, behavioral avoidance, exposure heterogeneity, residual activity, dose distribution, and frequency of use. Experience with spinosyn insecticides demonstrates that resistance to a naturally derived insecticide class can be inherited and can produce cross-resistance within related compounds [20]. The implication is not that all natural products share the same liability, but that novelty of origin cannot substitute for resistance-specific evidence.

Cross-resistance is particularly important when a candidate is proposed for populations already exposed to related targets or metabolic selection pressures. In resistant stored-product insects, changes in susceptibility can persist, decline, or interact with biochemical traits after selection, making resistance status a dynamic population property rather than a fixed label [21]. Candidate testing should therefore include susceptible and relevant resistant populations whenever plausible target-site or metabolic overlap exists. A candidate may retain activity against one resistant strain while failing against another because the underlying resistance mechanisms differ. Similarly, apparent absence of cross-resistance in a short-term assay does not demonstrate low evolutionary risk. It only indicates that the tested population, mechanism, exposure route, and endpoint did not reveal cross-resistance under those conditions.

Long-term evidence from spinosyn use shows that resistance and cross-resistance patterns are shaped by target changes, metabolism, inheritance, fitness effects, and deployment

history [22]. These lessons support a staged resistance assessment for natural-product candidates: baseline susceptibility should be established; existing resistant populations should be tested; plausible target-site and metabolic overlap should be examined; selection experiments should be conducted where justified; and resistance stability, inheritance, and fitness consequences should be characterized before durability claims are made. For chemically complex extracts, the contribution of each major active constituent should also be considered because a mixture can impose shared, sequential, or uneven selection pressures. Studies of botanically derived candidates against *Spodoptera frugiperda* illustrate the importance of connecting efficacy with biochemical and physiological responses rather than treating activity as evidence of resistance-breaking capacity [23]. A natural product should therefore be prioritized for resistance management only when its activity against resistant insects, mechanism profile, exposure pattern, and selection consequences are directly examined.

Proposed natural-product prioritization criteria

The proposed prioritization structure begins with intended-use definition and chemical identity. The intended pest, life stage, crop or stored-product setting, application route, required persistence, and principal non-target exposure pathways should be stated before comparative ranking. Candidate identity should then be established at the level appropriate to the proposed product: authenticated plant material, standardized extract, essential oil, fraction, isolated compound, derivative, or defined mixture. Activity that combines behavioral modification with toxicity may be valuable, but its contribution depends on whether repellency, attraction, feeding disruption, or mortality supports the intended control objective [24]. A behaviorally active candidate should not advance merely because it changes movement in a laboratory arena; the direction, duration, operational consequence, and possibility of pest redistribution must be determined.

The next stage integrates efficacy, mechanism, selectivity, interaction, stability, resistance, and formulation. Candidates should progress only when each domain is sufficiently characterized

for the intended decision, rather than when a single favorable property compensates silently for several unresolved liabilities. Defined plant-oil mixtures may offer useful activity against difficult pests, but their component ratios, delivery conditions, and biological interactions must remain reproducible [25]. Commercial development further requires scalable sourcing, analytical specifications, manufacturing consistency, storage stability, application compatibility, and a defensible use pattern [26].

These relations form a staged pathway rather than a universal numerical score: failure at an early identity or reproducibility gate can invalidate later comparisons, whereas uncertainty at a later gate may justify targeted investigation rather than immediate rejection.

Figure 1 presents a staged candidate-prioritization pathway from chemical discovery to insect-control relevance within the analytical logic developed in this section.

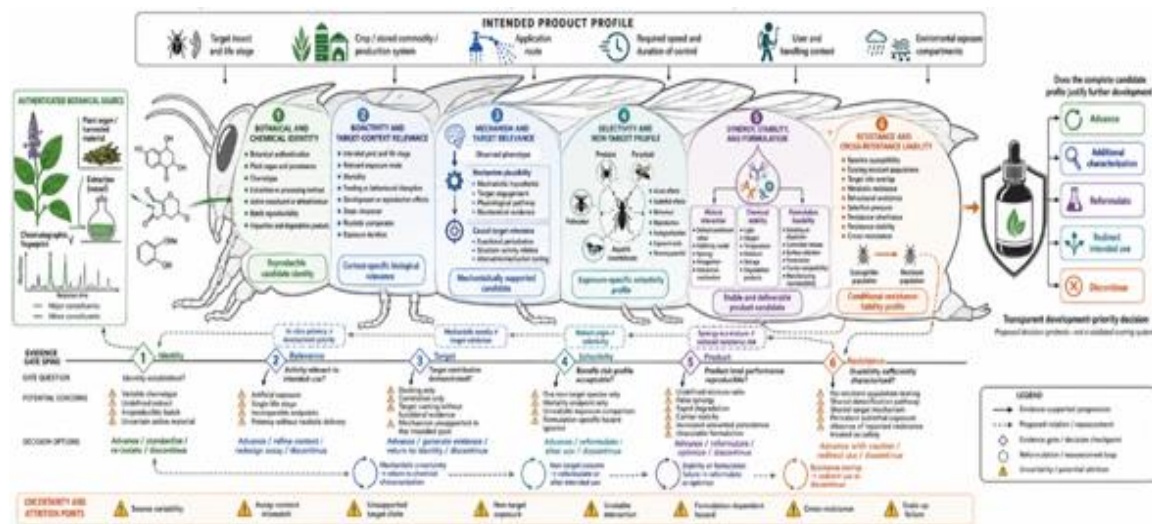


Figure 1. A staged candidate-prioritization pathway from chemical discovery to insect-control relevance

Alt text

A structured conceptual diagram that presents a staged candidate-prioritization pathway from chemical discovery to insect-control relevance, with labelled components, directional relations, contextual modifiers, uncertainty points, and a clear boundary between observed evidence and proposed synthesis.

The final stage is a transparent advancement decision. A candidate may advance, require reformulation, return for mechanism or safety clarification, be redirected to a different use, or be discontinued. The decision should be based on the remaining uncertainty that is most capable of changing development priority. Formulation strategies can improve delivery and stability, but they can also introduce new exposure pathways,

materials, manufacturing demands, and validation requirements [27]. Consequently, a candidate should not be described as development-ready unless the evidence applies to the complete product under the intended use conditions. The proposed structure is an organizing synthesis, not a validated framework, and it does not prescribe fixed thresholds or universal weights. Its purpose is to make the basis of prioritization explicit and to prevent potency, novelty, natural origin, or mixture complexity from functioning as unsupported proxies for overall merit.

The proposed components, evidence bases, boundary conditions, failure modes, and validation requirements are organized in **Table 2**.

Table 2. Proposed Natural-Product Prioritization Criteria: Components, Evidence Basis, Relations, Boundary Conditions, Failure Modes, and Validation Requirements

Proposed component	Purpose	Evidence basis	Relation or mechanism	Input or precondition	Expected output	Boundary condition or failure mode	Validation requirement
Intended-use definition	Anchor all subsequent comparisons to a	Target biology, production setting, exposure route,	Determines which efficacy, safety,	Defined pest, life stage, setting,	Intended product profile	Generic pesticidal activity may not	Prospective testing under conditions

	practical insect-control need	required duration, and user context	stability, and delivery evidence is relevant	application method, and desired outcome		fit the intended use	representing the defined use
Botanical and chemical identity	Establish what material is being evaluated	Taxonomic, analytical, and processing characterization	Links biological effects to a reproducible candidate	Authenticated source, defined extraction or isolation, and analytical profile	Traceable candidate specification	Chemotype, harvest, processing, or degradation variability can invalidate comparisons	Independent batch characterization and reproducibility testing
Bioactivity relevance	Determine whether activity supports the intended control objective	Mortality, development, reproduction, feeding, behavior, or other relevant endpoints	Connects exposure to a biologically and operationally meaningful response	Appropriate pest, life stage, route, comparator, and exposure duration	Context-specific efficacy profile	High potency in an artificial assay may not translate to practical control	Realistic laboratory, semi-field, storage, greenhouse, or field evaluation as appropriate
Mechanism plausibility	Explain how the candidate produces the observed effect	Physiological, biochemical, molecular, and pharmacological evidence	Narrows competing explanations and guides optimization	Chemically defined candidate and reproducible phenotype	Supported mechanistic hypothesis	Correlation, docking, or target naming may be mistaken for causation	Functional perturbation, target-engagement, rescue, or convergent mechanistic evidence
Target relevance	Establish whether the proposed target contributes causally in the intended pest	Target-specific functional evidence	Links molecular engagement to insecticidal phenotype	Plausible mechanism and accessible target	Target-supported candidate profile	Novel target description may lack causal or operational relevance	Genetic, biochemical, pharmacological, or structure–activity validation in the relevant system
Selectivity profile	Compare desired pest effects with plausible non-target effects	Target and non-target lethal, sublethal, behavioral, and functional endpoints	Integrates intrinsic sensitivity with exposure	Defined formulation and intended application pattern	Exposure-specific benefit–risk profile	Testing one non-target species or only acute mortality may create false reassurance	Tiered testing across relevant beneficial and environmental compartments
Mixture-interaction characterization	Determine whether combinations are additive, synergistic, or antagonistic	Defined-component and ratio-specific experiments	Components may alter penetration, metabolism, target action, volatility, or release	Known composition and individually tested constituents	Reproducible interaction profile	Undefined ratios or inappropriate additivity models can produce false synergy claims	Independent replication, ratio analysis, mechanistic follow-up, and formulated-mixture testing
Stability characterization	Determine whether active chemistry remains usable during storage and application	Chemical, physical, thermal, oxidative, and photolytic stability evidence	Governs retained active content and temporal exposure	Defined candidate, packaging, storage conditions, and use environment	Stability and degradation profile	Stabilization may increase unwanted environmental persistence	Product-level storage, release, degradation, and residue studies
Formulation feasibility	Deliver the candidate safely and effectively to the intended target	Solubility, dispersion, release, adhesion, penetration, compatibility, and manufacturability	Converts active chemistry into an applicable product	Defined active material and intended application method	Reproducible prototype formulation	Carrier toxicity, complexity, scale-up limitations, or altered exposure can outweigh efficacy gains	Comparative product testing, process reproducibility, and application-system compatibility
Resistance-liability assessment	Determine potential durability and fit with resistance management	Baseline susceptibility, mechanism overlap, selection, inheritance, and cross-resistance evidence	Links target and exposure pattern to evolutionary selection	Defined active profile and relevant insect populations	Conditional resistance-risk characterization	Absence of reported resistance may be mistaken for low risk	Resistant-population testing, selection studies, mechanistic analysis, and longitudinal monitoring
Integrated advancement decision	Determine whether development, reformulation, redirection, or discontinuation is justified	Combined chemical, biological, safety, formulation, and resistance evidence	Makes trade-offs, uncertainty, and evidence gaps explicit	Completed domain-specific assessment appropriate to the decision stage	Transparent advancement rationale	One favorable domain may obscure decisive uncertainty elsewhere	Prospective application of the criteria across diverse candidates and independent decision settings

Development and research implications

The first development priority is infrastructure for reproducible candidate definition. Botanical biopesticide research requires stronger links among plant authentication, chemotype documentation, cultivation or collection conditions, extraction, analytical profiling, formulation, and biological testing. Regional assessments of botanical-biopesticide development show that scientific potential coexists with limitations in standardization, product development, commercialization, and evidence transfer [28]. Progress would be demonstrated not by a larger number of active extracts, but by a larger proportion of candidates that can be reproduced across laboratories and batches using explicit chemical specifications. Shared reporting expectations should therefore include source identity, processing history, extraction yield, full or marker-based chemical profiles, storage conditions, formulation composition, and testing of independent batches. The second priority is comparative biological validation across target and non-target contexts. Candidate studies should move beyond single-species acute assays toward designs that compare relevant pest stages, resistant populations where appropriate, beneficial organisms, and environmental compartments under aligned exposure scenarios. Essential oils proposed against aphids illustrate both the breadth of available chemistry and the difficulty of translating laboratory activity into consistent pest-management value [29]. Progress would be indicated by reproducible performance across realistic exposure conditions, identification of the life stages and routes for which the candidate is most suitable, and demonstration that target control is compatible with biological-control organisms and other relevant non-targets. Negative or context-dependent findings should be reported because they clarify boundaries and prevent repeated investment in unsuitable use patterns.

The third priority is integration of mechanism, formulation, and resistance research. Molecular investigation can identify targets, detoxification responses, and structure-activity relations, while formulation research can determine whether relevant exposure can be achieved and maintained. These strands should be evaluated together because a validated target is of limited

development value if the compound cannot reach it under practical conditions, and an effective formulation is difficult to manage responsibly if its mechanism and resistance liabilities remain obscure. Molecular insight into plant-insect interactions can guide candidate optimization, but it must be linked to chemical reproducibility, ecological context, and product-level exposure [30]. Progress would be demonstrated by prospective studies in which chemically defined candidates pass through explicit evidence gates and by independent testing of whether the proposed prioritization structure improves the consistency, transparency, or predictive value of advancement decisions. Until such validation is performed, the structure should remain a scholarly decision aid rather than a deployment-ready framework.

CONCLUSION

Natural products merit development as insecticides when their promise extends beyond an isolated observation of bioactivity. The strongest candidates are chemically reproducible, relevant to a defined pest and use context, supported by appropriately bounded mechanistic evidence, selectively effective under plausible exposure, compatible with a feasible and stable formulation, and accompanied by a resistance assessment that considers existing and potential cross-resistance. No single domain is sufficient: *in vitro* potency is not equivalent to development priority, mechanistic novelty is not equivalent to target validation, natural origin is not equivalent to selectivity, and synergy in a mixture is not equivalent to reduced resistance risk. Candidate advancement should therefore proceed through transparent evidence gates that identify which uncertainties are tolerable, which require targeted resolution, and which undermine the proposed product concept. The highest-priority implication is to evaluate natural products as complete, intended-use-specific candidates rather than as chemically or biologically interesting substances in isolation. The proposed structure offers an explicitly non-validated basis for organizing such decisions and should be tested prospectively across different botanical materials, insect taxa, formulations, and production systems.

ACKNOWLEDGMENTS: None

CONFLICT OF INTEREST: None

FINANCIAL SUPPORT: None

ETHICS STATEMENT: None

REFERENCES

1. Isman MB. Botanical insecticides in the twenty-first century—fulfilling their promise? *Annu Rev Entomol.* 2020;65:233-49. doi:10.1146/annurev-ento-011019-025010
2. Marrone PG. Pesticidal natural products—status and future potential. *Pest Manag Sci.* 2019;75(9):2325-40. doi:10.1002/ps.5433
3. Campos EVR, Proença PLF, Oliveira JL, Bakshi M, Abhilash PC, Fraceto LF. Use of botanical insecticides for sustainable agriculture: Future perspectives. *Ecol Indic.* 2019;105:483-95. doi:10.1016/j.ecolind.2018.04.038
4. Stevenson PC, Isman MB, Belmain SR. Pesticidal plants in Africa: A global vision of new biological control products from local uses. *Ind Crops Prod.* 2017;110:2-9. doi:10.1016/j.indcrop.2017.08.034
5. Isman MB. Bridging the gap: Moving botanical insecticides from the laboratory to the farm. *Ind Crops Prod.* 2017;110:10-14. doi:10.1016/j.indcrop.2017.07.012
6. Haddi K, Turchen LM, Viteri Jumbo LO, Guedes RNC, Pereira EJJ, Aguiar RWS, et al. Rethinking biorational insecticides for pest management: Unintended effects and consequences. *Pest Manag Sci.* 2020;76(7):2286-93. doi:10.1002/ps.5837
7. Price CSV, Campbell H, Pope TW. Assessing the potential of biopesticides to control the cabbage stem flea beetle *Psylliodes chrysocephala*. *Pest Manag Sci.* 2024;80(5):2471-9. doi:10.1002/ps.7746
8. Li T, Zhang J, Ma S, Gao L, Chen C, Ji Z, et al. Identification and mechanism of insecticidal periplocosides from the root bark of *Periploca sepium* Bunge. *Pest Manag Sci.* 2021;77(4):1925-35. doi:10.1002/ps.6220
9. Huo X, Ma H, Zhu H, Liu J, Zhou Y, Zhou X, et al. Identification and pharmacological characterization of the voltage-gated potassium channel Shab in diamondback moth, *Plutella xylostella*. *Pest Manag Sci.* 2023;79(3):1251-60. doi:10.1002/ps.7300
10. Ding Y, Chen S, Zhang F, Li W, Ge G, Liu T, et al. Chitinase is a potent insecticidal molecular target of camptothecin and its derivatives. *J Agric Food Chem.* 2023;71(4):1845-51. doi:10.1021/acs.jafc.2c06607
11. Jesser E, Yeguerman C, Stefanazzi N, Gomez R, Murray AP, Ferrero AA, et al. Ecofriendly approach for the control of a common insect pest in the food industry, combining polymeric nanoparticles and post-application temperatures. *J Agric Food Chem.* 2020;68(21):5951-8. doi:10.1021/acs.jafc.9b06604
12. Novák M, Pavela R, Spinozzi E, Ferrati M, Petrelli R, Maggi F, et al. Lethal and sublethal effects of carlina oxide on the aphid *Metopolophium dirhodum* and its non-target impact on two biological control agents. *J Pest Sci.* 2024;97(4):2131-8. doi:10.1007/s10340-024-01768-z
13. Passos LC, Ricupero M, Gugliuzzo A, Soares MA, Desneux N, Campolo O, et al. Sublethal effects of plant essential oils toward the zoophytophagous mirid *Nesidiocoris tenuis*. *J Pest Sci.* 2022;95(4):1609-19. doi:10.1007/s10340-022-01548-7
14. Nederstigt TAP, Huijter VN, Planjer D, Eekhout FJM, Barmantlo SH, Peijnenburg WJGM, et al. What is there to gain and lose from plant-derived nano-enabled pesticides: Eugenol-loaded nanocarriers exert long-lived effects on non-target freshwater invertebrate communities. *J Hazard Mater.* 2025;496:139560. doi:10.1016/j.jhazmat.2025.139560
15. Pavela R, Morshedloo MR, Mumivand H, Khorsand GJ, Karami A, Maggi F, et al. Phenolic monoterpene-rich essential oils from Apiaceae and Lamiaceae species: Insecticidal activity and safety evaluation on non-target earthworms. *Entomol Gen.* 2020;40(4):421-35. doi:10.1127/entomologia/2020/1131
16. Jesser E, Yeguerman C, Gili V, Santillan G, Murray AP, Domini C, et al. Optimization and characterization of essential oil nanoemulsions using ultrasound for new ecofriendly insecticides. *ACS Sustain Chem*

- Eng. 2020;8(21):7981-92. doi:10.1021/acssuschemeng.0c02224
17. Kim S, Yoon J, Tak JH. Synergistic mechanism of insecticidal activity in basil and mandarin essential oils against the tobacco cutworm. *J Pest Sci.* 2021;94(4):1119-31. doi:10.1007/s10340-021-01345-8
 18. Giunti G, Campolo O, Laudani F, Zappalà L, Palmeri V. Bioactivity of essential oil-based nano-biopesticides toward *Rhyzopertha dominica* (Coleoptera: Bostrichidae). *Ind Crops Prod.* 2021;162:113257. doi:10.1016/j.indcrop.2021.113257
 19. Isman MB, Norris EJ. Bioinsecticide synergy: The good, the bad and the unknown. *Curr Opin Environ Sci Health.* 2024;42:100583. doi:10.1016/j.coesh.2024.100583
 20. Lira EC, Bolzan A, do Nascimento ARB, Amaral FSA, Kanno RH, Kaiser IS, et al. Resistance of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) to spinetoram: Inheritance and cross-resistance to spinosad. *Pest Manag Sci.* 2020;76(8):2674-80. doi:10.1002/ps.5812
 21. Wang YC, Chen YC, Chen CY, Chen ME. Resistance reduction and cross-resistance of spinosad-resistant *Rhyzopertha dominica* (Coleoptera: Bostrichidae) and the association between spinosad resistance and maltase activity. *Pest Manag Sci.* 2025;81(6):2840-6. doi:10.1002/ps.8650
 22. Sparks TC, Wessels FJ, Perry T, Price MJ, Siebert MW, Mann DGJ. Spinosyn resistance and cross-resistance—a 25 year review and analysis. *Pestic Biochem Physiol.* 2025;210:106363. doi:10.1016/j.pestbp.2025.106363
 23. Changke V, Nobsathian S, Le Goff G, Coustau C, Bullangpoti V. Insecticidal efficacy and possibility of *Combretum trifoliatum* Vent. (Myrtales: Combretaceae) extracts in controlling *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae). *Pest Manag Sci.* 2023;79(12):4868-78. doi:10.1002/ps.7688
 24. Ghabbari M, Guarino S, Caleca V, Saiano F, Sinacori M, Baser N, et al. Behavior-modifying and insecticidal effects of plant extracts on adults of *Ceratitis capitata* (Wiedemann) (Diptera Tephritidae). *J Pest Sci.* 2018;91(2):907-17. doi:10.1007/s10340-018-0952-6
 25. Abdelatti ZAS, Hartbauer M. Plant oil mixtures as a novel botanical pesticide to control gregarious locusts. *J Pest Sci.* 2020;93(1):341-53. doi:10.1007/s10340-019-01169-7
 26. Isman MB. Commercial development of plant essential oils and their constituents as active ingredients in bioinsecticides. *Phytochem Rev.* 2020;19(2):235-41. doi:10.1007/s11101-019-09653-9
 27. Sarmah K, Anbalagan T, Marimuthu M, Mariappan P, Angappan S, Vaithiyanathan S. Innovative formulation strategies for botanical- and essential oil-based insecticides. *J Pest Sci.* 2025;98(1):1-30. doi:10.1007/s10340-024-01846-2
 28. Acheuk F, Basiouni S, Shehata AA, Dick K, Hajri H, Lasram S, et al. Status and prospects of botanical biopesticides in Europe and Mediterranean countries. *Biomolecules.* 2022;12(2):311. doi:10.3390/biom12020311
 29. Ikbali C, Pavela R. Essential oils as active ingredients of botanical insecticides against aphids. *J Pest Sci.* 2019;92(3):971-86. doi:10.1007/s10340-019-01089-6
 30. Fuentes-Lopez K, Olivero-Verbel J, Caballero-Gallardo K. Aromatic plant-insect dynamics: Molecular insights for advancing pest control strategies. *J Pest Sci.* 2025;98(4):2501-17. doi:10.1007/s10340-025-01953-8