



## Food Baits and Traps for Monitoring *Drosophila Suzukii* and *Zaprionus Indianus* “Drosophilidae” in Fig Orchards

Liliana Arios-Caro<sup>1</sup>, Víctor López-Martínez<sup>1\*</sup>, Iran Alia-Tejacal<sup>1</sup>, Dagoberto Guillén-Sánchez<sup>2</sup>, Porfirio Juárez-López<sup>1</sup>, Nidia Bélgica Pérez-De la O<sup>3</sup>

<sup>1</sup>Faculty of Agricultural Sciences, Autonomous University of the State of Morelos, Cuernavaca, Mexico.

<sup>2</sup>Xalostoc School of Higher Studies, Autonomous University of the State of Morelos, Cuernavaca, Mexico.

<sup>3</sup>Phytosanitary-Entomology and Acarology Postgraduate Program, College of Postgraduate, Montecillo, Mexico.

### ABSTRACT

*Drosophila suzukii* Matsumura and *Zaprionus indianus* Gupta (Diptera: Drosophilidae) are invasive pests that attack fig crops in Mexico. Monitoring fruit flies' populations' density is the main tool for taking control action choices. In the case of fruit flies, the use of food baits and traps is a regular practice in fruit and vegetable production. However, local fig growers are characterized by a low technological level, with a constant search for economical and practical options to improve their quality and incomes. Here we tested three traps (two from formal manufacturers and one plastic handmade) and three food baits for collecting adults of *D. suzukii* and *Z. indianus*. Two trials were conducted in different periods, November-December 2018 and July-August 2019 in local fig orchards. Handmade plastic jar traps baited with commercial food baits showed good performance against commercial traps, for the two fruitflies in both evaluation periods. The role of the handmade trap design in the attraction efficiency for fruitflies collection is discussed. The region of study is considered with a high population level of *D. suzukii* and *Z. indianus*.

**Keywords:** Fig flies, Drosophilid flies, Trapping, Handmade trap.

**HOW TO CITE THIS ARTICLE:** Arios-Caro L, López-Martínez V, Alia-Tejacal I, Guillén-Sánchez D, Juárez-López P, Pérez-De la O NB. Food Baits and Traps for Monitoring *Drosophila Suzukii* and *Zaprionus Indianus* “Drosophilidae” in Fig Orchards. *Entomol Appl Sci Lett.* 2021;8(4):43-51. <https://doi.org/10.51847/gDQOwilx49>

**Corresponding author:** Víctor López-Martínez

**E-mail** ✉ [victor.lopez@uaem.mx](mailto:victor.lopez@uaem.mx)

**Received:** 25/08/2021

**Accepted:** 19/12/2021

### INTRODUCTION

The fig tree (*Ficus carica* L., Moraceae) is a native species from the south-western Mediterranean area and is considered the first plant domesticated by human beings [1]. As a crop, is cultivated in temperate, tropical, and subtropical regions, its rusticity has allowed being grown in great soils diversity [2]. Worldwide, the cultivated area exceeds 376,100 hectares, with an estimated production of 1,064,400 tons; Turkey is considered as the main fig producer [3]. In Mexico, the cultivated surface reported in 2018 was 1,357 ha, a production volume of 7,700

tons, and an economic value of 8 million dollars [4]. The most important production areas are concentrated in the central Morelos state, characterized by their low technological level [5]. The principal tool for pest management is the periodical chemical pesticides application. Recently, two Drosophilidae (Insecta: Diptera) have been affecting quality and phytosanitary fruit status: *Drosophila suzukii* Matsumura and *Zaprionus indianus* Gupta [6, 7], considered devastating pests of strawberry, peach, blackberries, figs, and small fruits with thin epicarp [5, 8, 9].

*Drosophila suzukii* is original from southeast Asia [10], described from Japan in 1931 [11]. In Mexico, *D. suzukii* was reported as an invasive species in 2011 and was considered as present in only one municipality: Los Reyes (Michoacan) [12]. Is presently considered as Mexico and under governmental control, although only in some areas with cultivated hosts meets the formal quarantine pest definition [7, 13]. Unlike other drosophilid flies, *D. suzukii* is characterized by a serrated ovipositor, used to easily cut the fruit epicarp to oviposit into ripened and unripened fruits; this species feeds on overripe or decaying fruits [14, 15].

*Zaprionus indianus* was reported for the first time in America in fig fruits from Brazil, causing losses of 40 a 50% in commercial figs, and where it was called the Africa fig fly [16]. It was detected in Mexico for the first time in Chiapas in 2002 [17], despite the fact it has received little attention, its distribution includes crops from seven federal entities [6, 7, 17]. In fig is considered a major pest [18]. *Zaprionus indianus* unlike *D. suzukii* does not have a serrated ovipositor, this fly penetrates the ostiole to oviposit when the fig is reaching its maturity [19].

As part of the strategies implemented for these drosophilid management, trapping for monitoring adult populations in fig production areas and regions with invasion risk has been implemented [20]. Monitoring with traps allows to determine the drosophilid adult presence or absence and to calculate population fluctuations. Traps used by local growers are plastic hand man jars, baited with apple vinegar, disposed of in quadrants separated at 1 km [5] but mainly the typical Multilure® baited for other fruitflies (personal observation). Different types of commercial and homemade baits based on apple cider vinegar, wine, and yeast have been evaluated for the monitoring of these flies [21]. However, apple vinegar has the potential to attract a wide diversity of Drosophilidae flies, included not target species, and other Diptera, Lepidoptera, Hymenoptera, and Coleoptera species [22, 23].

There are commercial traps (Multilure® and Pherocon SWD®) and food attractants (Pherocon SWD® Dual-Lure, Suzukii Trap®) recommended for monitoring and even for mass trapping for these and other fruitflies, but they never have been tested under local grower

conditions in Morelos. The purpose of this work was to evaluate the efficacy of commercial baits and traps for monitoring adults of two drosophilid adults (Insecta: Diptera) with economic importance for cultivated fig in Morelos, Mexico.

## MATERIALS AND METHODS

### *Experiment location*

The research was carried out in commercial fig orchards, variety Black Mission, grown in Ayala, Morelos, Mexico (18.734206° -98.915858). The climate in the region is warm subhumid with an average annual temperature of 24°C [24].

### *Traps and food baits*

Two commercial, Multilure® (Ferommis, Mexico) and Pherocon SWD® (Trécé, Inc., United States) and one handmade jar trap were used. Handmade traps were made with one-liter plastic jars, with 10 holes of 4 mm around the upper container, and a red base [5, 25]. As food baits, Suzukii Trap® (Bioiberica, Spain), Pherocon SWD® Dual-Lure® (Ferommis, Mexico), and Cera Trap® (Agrotecnologia Alternativa, S.A. de C.V., Mexico) were analyzed. 250 ml of liquid baits (Suzukii Trap® and Cera Trap®) was placed by trap, for Pherocon SWD® Dual-Lure, one dispenser was attached in the interior of the traps, and soapy water (5%) was used for insect retention.

### *Experimental design*

Two factors were analyzed, trap (with three levels) and food bait (three levels), combination of both factors produced nine treatments 1) Handmade plastic jar-Cera trap®; 2) Handmade plastic jar-Suzukii Trap®; 3) Handmade plastic jar-Pherocon SWD® Dual-Lure®; 4) Multilure®- Suzukii Trap®; 5) Multilure®- Suzukii Trap®; 6) Multilure®- Pherocon SWD® Dual-Lure®; 7) Pherocon®- Suzukii Trap®; 8) Pherocon®- Suzukii Trap®; and 9) Pherocon®- Pherocon SWD® Dual-Lure®.

The experimental design was completely randomized in blocks, each treatment had four repetitions, and all treatments were arranged in four rows. Traps were checked weekly, and at each sampling date, treatments were randomized to avoid any positional bias. The separation distance between traps was 20 m and between rows was 30 m, traps were placed at  $\frac{3}{4}$

height of trees, avoiding direct exposure to the sun. The experiment was carried out on two occasions, with a duration of six weeks. The first period was from 2018 November 19<sup>th</sup> to December 24<sup>th</sup>, and the second from 2019 July 12<sup>th</sup> to August 16<sup>th</sup>. On both dates, fruits were in  $\frac{3}{4}$  of their physiological maturity. Pherocom SWD® Dual-Lure© dispenser was changed every four weeks, and the soapy water weekly. Suzukii trap® and Cera trap® baits were filled when necessary.

#### Entomological determination

All specimens captured were preserved in alcohol 70% and determined with taxonomic keys [26].

#### Data analysis

Data was tested for normality and homogeneity of variance by species, then treatment means were compared (ANOVA), and when significant differences were detected the Fischer test was applied ( $\alpha=0.05$ ). In the second period, data was transformed ( $\sqrt{X+0.5}$ ) to normalize the distribution and homogenize the variations before analysis.

#### Day Trap Flies index (DTF)

The Day Trap Flies index is used to calculate the relative fruitflies adult presence in a particular area and time, commonly used in native fruit flies of the genus *Anastrepha* (Diptera: Tephritidae), used to choose management strategies for tephritid populations [27]. The index was calculated by species by treatment [5]:

$$DTF = F/(TxD) \quad (1)$$

Donde:

F= Number of flies collected

T = Total number of traps revised

D = Number of days those traps were exposed in the field

DTF is expressed in the format 0.0000 [28], level prevalence is categorized as high ( $\geq 0.0100$ ), low ( $\leq 0.0100$ ), and absent (0.0000).

## RESULTS AND DISCUSSION

Monitoring is an important tool to determine and know the present/absence of pests in a crop and allows making correct decisions for their integrated management [21]. For fruit flies, trap design and food baits influence capturing adults [29, 30]. The trap design must be attractive for fruit flies, with a capacity for temporal storage of the food bait, and with space for the release of the bait compounds; at the same time, food bait needs to be the correct lure and could act as the retention mechanism [31].

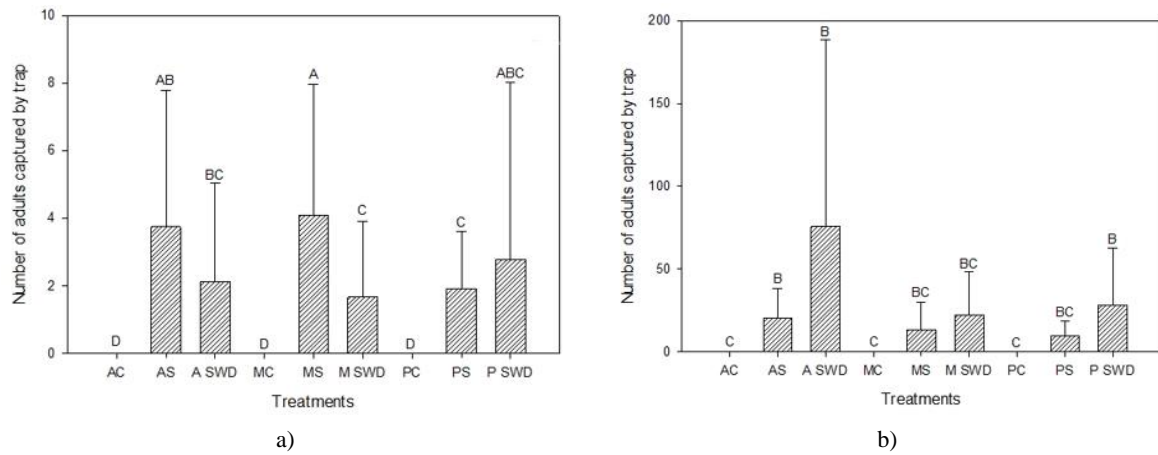
#### First trial

##### Traps and food baits

4,458 specimens were captured, 8.0 % (373) were determined as *D. suzukii* and 92% (4,285) were classified as *Z. indianus*. The type of food bait affected the drosophilid species captured. Suzukii trap© was more effective to capture *D. suzukii*, but only when is used in combination with the handmade or Multilure® traps; its attraction is reduced with Pherocom© trap (**Figure 1a**). However, this food bait captured not target Drosophilidae (data not shown). Pherocom© trap plus Pherocom SWD© Dual-Lure© food bait showed similar captures in comparison with Suzukii trap© in handmade or Multilure® traps.

For *Z. indianus*, handmade plastic traps baited with Pherocom SWD© were the best combination, collecting three or five times the rest of the treatments (**Figure 1b**).

For both drosophilid species, Cera Trap® did not capture specimens, independently of the trap used (**Figures 1a and 1b**).



**Figure 1.** *Drosophila suzukii* (a) and *Zapronius indianus* (b) captures in a combination of three traps and food baits in ficus, Mexico (November-December 2018). AC: Handmade trap + Cera Trap©; AS: Handmade trap+ Suzukii© trap; ASWD: Handmade trap + Pheroacom SWD© Dual-Lure©; MC: Multilure© trap + Cera Trap©; MS: Multilure© trap + Suzukii© trap; MSWD: Multilure© trap + Pheroacom SWD© Dual-Lure©; PC: Pheroacom© trap + Cera Trap©; PS: Pheroacom© trap + Suzukii© trap; PSWD: Pheroacom© trap + Pheroacom SWD© Dual-Lure©.

*DTF index*

Interaction between traps and food baits affected the prevalence population-level calculated for *D. suzukii*: the biggest DTF index value was calculated with Multilure© trap and Suzukii trap© as food bait and similar when this food bait is used with handmade plastic jar traps. Pheroacom© trap plus Pheroacom SWD© Dual-Lure© bait had a similar value as Multilure© trap

and Suzukii trap© combination (**Table 1**). The rest of the treatments showed minor sensibility to calculate the DTF index, and even those traps baited with Cera trap© did not allow to calculate it. Food baits Suzukii trap© and Pheroacom SWD© Dual-Lure© used with all three traps calculate a high population prevalence for *D. suzukii*.

**Table 1.** Day Trap Flies (DTF) population prevalence index, maximum and minimum values calculated for *Drosophila suzukii* with different traps and food baits in figs from Morelos, Mexico (November-December 2018).

Treatment (trap plus food bait)	Maximum prevalence level	Minimum prevalence level	Weeks with prevalence values 0.0000
Handmade + Cera trap©	0.0000	0.0000	6
Handmade + Suzukii trap©	0.9200	0.2500	0
Handmade + Pheroacom SWD© Dual-Lure©	0.8500	0.0700	0
Multilure© + Cera trap©	0.0000	0.0000	6
Multilure© + Suzukii trap©	1.1700	0.3200	0
Multilure© + Pheroacom SWD© Dual-Lure©	0.5700	0.0300	0
Pheroacom© + Cera trap©	0.0000	0.0000	6
Pheroacom© + Suzukii trap©	0.4600	0.1700	0
Pheroacom© + Pheroacom SWD© Dual-Lure©	1.0000	0.0700	0

**Table 2.** Day Trap Flies (DTF) population prevalence index, maximum and minimum values calculated for *Zapronius indianus* with different traps and food baits in figs from Morelos, Mexico (November-December 2018).

Treatment (trap plus food bait)	Maximum prevalence level	Minimum prevalence level	Weeks with prevalence values 0.0000
Handmade + Cera trap©	0.0000	0.0000	6
Handmade + Suzukii trap©	8.8500	1.0700	0
Handmade + Pheroacom SWD© Dual-Lure©	33.4000	2.2800	0
Multilure© + Cera trap©	0.0000	0.0000	6
Multilure© + Suzukii trap©	2.8900	0.1740	0
Multilure© + Pheroacom SWD© Dual-Lure©	7.4200	1.2800	0

Pherocom® + Cera trap®	0.0000	0.0000	6
Pherocom® + Suzukii trap®	2.3500	0.7500	0
Pherocom® + Pherocom SWD® Dual-Lure®	8.8500	0.7800	0

A similar response for the DTF index was recorded for *Z. indianus*, high population prevalence was calculated with the food baits Suzukii trap® and Pherocom SWD® Dual-Lure®, in all the traps tested (Table 2). With the higher DTF index calculated by Handmade plastic jar handmade plus Pherocom SWD® Dual-Lure® bait. Cera Trap® as a food bait did not allow the calculation of DTF index independently of the trap analyzed (Table 2).

### Second trial

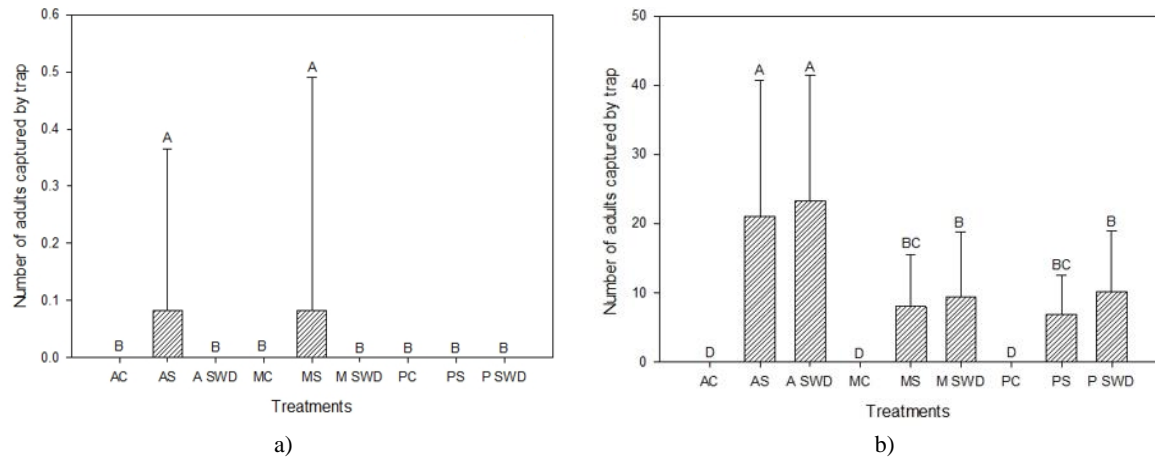
#### Traps and food baits

In the second trial, captures for both species were lower than the first trial. 1,899 were collected,

just four of *D. suzukii* (0.2 %), the rest of the specimens were determined as *Z. indianus* (1,895). All *D. suzukii* individuals were captured in the handmade plastic jar or Multilure® traps baited with Suzukii® trap, without statistical differences (Figure 2a).

In *Z. indianus*, type of trap affected the number of specimens captured, handmade plastic jar trap baited with Suzukii trap® or Pherocom SWD® Dual-Lure® collected more than 100% in comparison with the rest of treatments (Figure 2b).

Similar to the recorded in the first trial, Cera Trap® food bait did not capture specimens.



**Figure 2.** *Drosophila suzukii* (A) and *Zaprionus indianus* (B) captures in a combination of three traps and food baits in ficus, Mexico (July-August 2019). AC: Handmade trap + Cera Trap®; AS: Handmade trap+ Suzukii® trap; ASWD: Handmade trap + Pherocom SWD® Dual-Lure®; MC: Multilure® trap + Cera Trap®; MS: Multilure® trap + Suzukii® trap; MSWD: Multilure® trap + Pherocom SWD® Dual-Lure®; PC: Pherocom® trap + Cera Trap®; PS: Pherocom® trap + Suzukii® trap; PSWD: Pherocom® trap + Pherocom SWD® Dual-Lure®.

The performance achieved by the handmade plastic jar trap is probably influenced by the combination of two factors: perforations and color. The number and diameter of the holes (4 mm) around the plastic jar walls facility dispersion of the food bait to the environment and the same time the entry of the fruitflies. In contrast, the Pherocom® trap has two big lateral holes covered with a plastic mesh with a 2 mm diameter opening, which is possibly difficult for direct entry to the trap. Finally, the Multilure® trap is characterized by a wide-open mouth at the bottom and a yellow base; is possible that hole dimension allows flies easy entry, but at the same

time may also facilitate its exit if the retention media is not adequate. As was pointed before [32], increasing the entry trap areas decrease their capture capacity and its performance [29]. An additional issue is the possibility to increase captures of not *Drosophilidae* target, even *Diptera* or another bigger arthropod [32]. Handmade plastic jar traps showed the minor entry space, and this could be their first advantage in comparison to the other traps, being more selective to *drosophilid* flies [29]. Regarding the color, although it has been considered that red is attractive for the capture of *D. suzukii* adults [29], coinciding partially with

the handmade red base trap, some authors consider that other colors could generate a greater attraction response [33, 34]. It seems that more options for this variable still need to be explored.

The drosophilid species responded differently to the bait tested, *D. suzukii* was attracted mainly to Suzukii trap® and *Z. indianus* to Pherocom® SWD Dual-Lure®. *Drosophila suzukii* is attracted mainly by fermented substances such as wine and vinegar, as well as some yeasts [29]. Suzukii trap® is composed of enriched protein substances, with a good attraction response of *D. suzukii*, proposed to be used in mass trapping for this fruitfly [35]. On the other side, *Z. indianus* has a preference for juices, vinegar, and wines [34, 36], with a good response to Pherocom® SWD Dual-Lure® [37] because is composed of wine and vinegar volatile substances (acetic acid, ethanol, acetone, and methanol) [38]. The null response of both drosophila to Cera trap ® is related to its origin. It was designed and tested

for *Ceratitis capitata* Wiedeman, however is now widely used for many *Anastrepha* species [39]. Results derived from this work suggests the use of Suzukii trap® and Pherocom® SWD Dual-Lure®, but Cera trap ® must be discarded immediately as bait food for these two drosophilid flies. The optimal selection of trap and food bait for fruiflies is a continuous task [40], as new designs and baits are being offered constantly, the options for local growers will increase their capacity for deploying better trapping systems.

#### DTF index

The population density in the second trial affected calculating the DTF index (**n n**). Just two treatments can calculate the index, Handmade plastic jar plus Suzukii trap® and Multilure® trap plus Suzukii trap® bait, with a high prevalence level (0.0300-0.0700). According to the rest of the treatments, *D. suzukii* is absent in the study region in this period (**Table 3**).

**Table 3.** Day Trap Flies (DTF) population prevalence index, maximum and minimum values calculated for *Drosophila suzukii* with different traps and food baits in figs from Morelos, Mexico (July-August 2019).

Treatment (trap plus food bait)	Maximum prevalence level	Minimum prevalence level	Weeks with prevalence values 0.0000
Handmade + Cera trap®	0.0000	0.0000	6
Handmade + Suzukii trap®	0.0300	0.0000	4
Handmade + Pherocom SWD® Dual-Lure®	0.0000	0.0000	6
Multilure® + Cera trap®	0.0000	0.0000	6
Multilure® + Suzukii trap®	0.0700	0.0000	5
Multilure® + Pherocom SWD® Dual-Lure®	0.0000	0.0000	6
Pherocom® + Cera trap®	0.0000	0.0000	6
Pherocom® + Suzukii trap®	0.0000	0.0000	6
Pherocom® + Pherocom SWD® Dual-Lure®	0.0000	0.0000	6

**Table 4.** Day Trap Flies (DTF) population prevalence index, maximum and minimum values calculated for *Zaprionus indianus* with different traps and food baits in figs from Morelos, Mexico (July-August 2019).

Treatment (trap plus food bait)	Maximum prevalence level	Minimum prevalence level	Weeks with prevalence values 0.0000
Handmade + Cera trap®	0.0000	0.0000	6
Handmade + Suzukii trap®	5.4600	1.8200	0
Handmade + Pherocom SWD® Dual-Lure®	5.3900	1.0350	0
Multilure® + Cera trap®	0.0000	0.0000	6
Multilure® + Suzukii trap®	1.6700	0.4600	0
Multilure® + Pherocom SWD® Dual-Lure®	2.2500	0.8500	0
Pherocom® + Cera trap®	0.0000	0.0000	6
Pherocom® + Suzukii trap®	1.7800	0.4600	0
Pherocom® + Pherocom SWD® Dual-Lure®	2.4600	0.0000	1

A high prevalence level was calculated for *Z. indianus* in all the traps and food baited used,

except when Cera Trap® was involved. Handmade jar trap showed the higher index

values, with similar results using Suzukii trap® or Pherocom SWD® Dual-Lure® (Table 4). In this trap, the DTF index was two or three times higher than other treatments.

DTF index pointed for a high prevalence level for *D. suzukii* and *Z. indianus*, confirming that fig production area has the environmental conditions for the continuous reproduction of this species [7] and calling for the impulse of regional and coordinate activities to reduce populations and potential damage from this fruitflies.

### CONCLUSION

*Drosophila suzukii* Matsumura and *Zaprionus indianus* Gupta, are two fruit flies of economic importance for fig production in Morelos, Mexico, where environmental conditions provide an opportunity for their population to increase. For the optimal implementation of a monitoring program for drosophild adults, results derived from this study indicate that *D. suzukii* and *Z. indianus* could be trapped deploying handmade plastic jar traps baited with Suzukii trap® or Pherocom® SWD Dual-Lure®.

**ACKNOWLEDGMENTS:** None

**CONFLICT OF INTEREST:** None

**FINANCIAL SUPPORT:** Liliana Arios Caro received a graduate grant from CONACYT (fellowship 702226).

**ETHICS STATEMENT:** None

### REFERENCES

1. Kisley ME, Hartmann A, Bar-Yosef O. Early domesticated fig in the Jordan Valley. *Science*. 2006;312(5778):1372-4. doi:10.1126/science.1125910.
2. González AM, Grajal MJ. Higueras de Canarias Caracterización morfológica de variedades. *Islas Canarias, España: Instituto Canario de Investigaciones Agrarias*; 2011. 126p.
3. FAO. FAOSTAT. Roma, Italia: FAO; 2013 [cited 2018 Oct 28]. Available from: <http://faostat.fao.org>
4. SIAP. Anuario Estadístico de la Producción Agrícola. Ciudad de México, México: SIAP; 2018 [cited 2018 Dec 2]. Available from: [http://nube.siap.gob.mx/cierre\\_agricola/](http://nube.siap.gob.mx/cierre_agricola/)
5. Senasica. Campaña Nacional contra Moscas nativas de la fruta. Ciudad de México, México: SENASICA; 2016 [cited 2018 Oct 28]. Available from: <http://www.senasica.gob.mx/?id=4605>
6. Bautista N, Illescas CP, Lopez E, Velazquez LJ, Garcia CJ. Presence of Drosophilidae (Diptera: Ephydroidea) flies associated with fig fruits in Morelos, Mexico. *Fla Entomol*. 2017;100(4):813-6. doi:10.1653/024.100.0409
7. Domínguez S, López V, Martínez A, Delgadillo Á, Guillén D, Campos M. Dinámica poblacional de *Zaprionus indianus* y *Drosophila suzukii* en higo en el estado de Morelos, México. *Southwest Entomol*. 2021;45(4):1161-4. doi:10.3958/059.045.0432
8. Burrack HJ, Fernández GE, Spivey T, Kraus DA. Variation in selection and utilization of host crops in the field and laboratory by *Drosophila suzukii* Matsumura (Diptera: Drosophilidae), an invasive frugivore. *Pest Manag Sci*. 2013;69(10):1173-80. doi:10.1002/ps.3489
9. Asplen MK, Anfora G, Biondi A, Choi DS, Chu D, Daane, KM, et al. Invasion biology of spotted wing *Drosophila* (*Drosophila suzukii*): a global perspective and future priorities. *J Pest Sci*. 2015;88(3):469-94. doi:10.1007/s10340-015-0681-z
10. Oregon State University. A New pest attacking healthy ripening fruit in Oregon. Oregon, USA: Oregon State University; 2009 [updated 2009 October; cited 2019 May 2]. Available from: <https://extension.oregonstate.edu/pests-weeds-diseases/insects/new-pest-attacking-healthy-ripening-fruit-oregon-spotted-wing>
11. Sarto V, Sorribas R. *Drosophila suzukii* (Matsumura, 1931), nueva amenaza para las producciones agrícolas. *Phytoma*. 2011;234:54-9. doi:10.13140/2.1.4429.5049
12. NAPPO. Detection of spotted-winged *drosophila* (*Drosophila suzukii* Matsumura) in the Municipality of Los Reyes, State of Michoacan, Mexico 2011 [Internet]. Raleigh, USA: NAPPO; 2011 [updated 2011

- November 30; cited 2016 Mar 15] Available from: <https://www.pestalerts.org/official-pest-report/detection-spotted-winged-drosophila-drosophila-suzukii-matsumura-municipality>
13. Garcia CJ, Bravo D, Ruiz I, Romero G, Quezada A, Hernandez S, et al. Presencia de la mosca del vinagre de alas manchadas *Drosophila suzukii* (Mtsamura 1931) (Diptera: Drosophilidae) en Mexico. Entomol Agri. 2016;3:354-60.
  14. Atallah J, Teixeira L, Salazar R, Zaragoza G, Artyom K. The making of a pest: the evolution of a fruit-penetrating ovipositor in *Drosophila suzukii* and related species. Proc R Soc B. 2014;281(1781):20132840. doi:10.1098/rspb.2013.2840
  15. Muslimah NA, Hassan SM, Hassan A. Molecular Method VS. Traditional Methods for Estimating the Prevalence of Larval Trematode Infections in Some Red Sea Snails. Int J Pharm Res Allied Sci. 2019;8(2):190-7.
  16. Vilela CR. Is *Zaprionus indianus* Gupta, 1970 (Diptera, Drosophilidae) currently colonizing the Neotropical Region. Drosoph Inf Serv. 1999;82:37-9.
  17. Castrezana S. New records of *Zaprionus indianus* Gupta, 1970 (Diptera, Drosophilidae) in North America and a key to identify some *Zaprionus* species deposited in the *Drosophila* Tucson Stock Center. Dros Inf Serv. 2007;90:34-6.
  18. Vilela CR, Goñi B. Mosca-africana-do-figo, *Zaprionus indianus* (Diptera: Drosophilidae). 2nd edition. São Paulo, Brazil: Holos Editora; 2015.173p.
  19. Joshi NK, Biddinger DJ, Demchak K, Deppen A. First report of *Zaprionus indianus* (Diptera: Drosophilidae) in commercial fruits and vegetables in Pennsylvania. J Insect Sci. 2014;14(1):259. doi:10.1093/jisesa/ieu121
  20. SENASICA. Mosca del vinagre de alas manchadas (*Drosophila suzukii* Mastsumura). Ciudad de México, México: SENASICA; 2015 [cited 2019 January 7]. Available from: <https://www.gob.mx/senasica/documentos/mosca-del-vinagre-de-alas-manchadas>
  21. Harmon DS, Haseeb M, Kanga LH, Liburd OE. Evaluation of monitoring traps and lures for *Drosophila suzukii* (Diptera: Drosophilidae) in berry plantings in Florida. Insects. 2019;10(10):313. doi:10.3390/insects10100313
  22. Zhu J, Park KC, Baker TC. Identification of odors from overripe mango that attract vinegar flies, *Drosophila melanogaster*. J Chem Ecol. 2003;29(4):899-909. doi:10.1023/a:1022931816351.
  23. Qian K, Zhu JJ, Sims SR, Taylor DB, Zeng X. Identification of volatile compounds from a food-grade vinegar attractive to house flies (Diptera: Muscidae). J Econ Entomol. 2013;106(2):979-87. doi:10.1603/EC12424
  24. INIFAP. Sistema de alerta fitosanitaria del estado de Morelos. Ciudad de México, México: INIFAP; 2017 [cited 2018 December 2]. Available from: <http://siafemor.inifap.gob.mx>
  25. Cardenas JA, Chavero J. Manual para el manejo fitosanitario de la mosca del vinagre de las alas manchadas (*Drosophila suzukii* Matsumura). Ciudad de México, México: SAGARPA; 2014. 18p.
  26. Miller ME, Marshall SA, Grimaldi DA. A review of the species of *Drosophila* (Diptera: Drosophilidae) and genera of Drosophilidae of Northeastern North America. Can J Arthropod Identify. 2017;31:1-282. doi:10.3752/cjai.2017.31
  27. Sosa JM, López V, Villegas ÓG, Juárez P, Burgos A. Dinámica Poblacional de Moscas de la Fruta en Quintana Roo, México. Southwest Entomol. 2017;42(1):275-82. doi:10.3958/059.042.0124
  28. Barclay HJ, Enkerlin WR, Manoukis NC, Reyes J. Guidelines for the Use of Mathematics in Operational Area-Wide Integrated Pest Management Programmes Using the Sterile Insect Technique with a Special Focus on Tephritid Fruit Flies. Rome, Italy: FAO/IAEA; 2016. 95p.
  29. Lee JC, Burrack HJ, Barrantes LD, Beers EH, Dreves AJ, Hamby KA, et al. Evaluation of Monitoring Traps for *Drosophila suzukii* (Diptera: Drosophilidae) in North America. J Econ Entomol. 2012;105(4):1350-7. doi:10.1603/EC12132
  30. Lasa R, Velázquez OE, Ortega R, Acosta E. Efficacy of commercial traps and food odor attractants for mass trapping of *Anastrepha ludens* (Diptera: Tephritidae). J Econ



- Entomol. 2014;107(1):198-205.  
doi:10.1603/EC13043
31. Thomas DB, Holler TC, Heath RR, Salinas EJ, Moses AL. Trap-lure combinations for surveillance of *Anastrepha* fruit flies (Diptera: Tephritidae). *Fla Entomol.* 2001;84(3):344-51. doi:10.2307/3496491
  32. Renkema JM, Buitenhuis R, Hallet RH. Optimizing trap design and trapping protocols for *Drosophila suzukii* (Diptera: Drosophilidae). *J Econ Entomol.* 2014;107(6):2107-18. doi:10.1603/EC14254.
  33. Little CM, Rizzato AR, Charbonneau L, Chapman T, Hillier NK. Color preference of the spotted *Drosophila*, *Drosophila suzukii*. *Sci Rep.* 2019;9:16051. doi:10.1038/s41598-019-52425-w
  34. Lasa R, Gschaedler AC, Bello G, Williams T. Laboratory evaluation of trap color and vinegar, yeast and fruit juice lure combinations for monitoring of *Zaprionus indianus* (Diptera: Drosophilidae). *Int J Pest Manag.* 2020;66(3):279-87. doi:10.1080/09670874.2019.1636328
  35. Botta A, Carrión M. *Suzukii* trap: atrayente específico para la captura de *Drosophila suzukii*. *Phytoma.* 2013;253:39-40.
  36. Epsky ND, Gill MA. Laboratory and field age of aqueous grape juice bait and capture of *Zaprionus indianus* (Diptera: Drosophilidae). *J Econ Entomol.* 2017;110(3):1326-32. doi:10.1093/jee/tox057
  37. Joshi NK, Butler B, Demchack K, Biddinder D. Seasonal occurrence of spotted wing *drosophila* in various small fruits and berries in Pennsylvania and Maryland. *J Appl Entomol.* 2016;141(1-2):156-60. doi:10.1111/jen.12325
  38. Escudero LA. Estudios desarrollados sobre los métodos biotecnológicos disponibles para el seguimiento y control de *Drosophila suzukii* en España. *Phytoma.* 2015;269:20-4.
  39. Lasa R, Cruz A. Efficacy of new commercial traps and the lure Ceratrap® against *Anastrepha obliqua* (Diptera: Tephritidae). *Fla. Entomol.* 2014;97(4):1369-77. doi:10.1653/024.097.0411
  40. Bali EM, Moraiti CA, Ioannou CS, Mavraganis V, Papadopoulus NT. Evaluation of mass trapping devices for early seasonal management of *Ceratitis capitata* (Diptera: Tephritidae) populations. *Agronomy.* 2021;11(6):1101. doi:10.3390/agronomy11061101