



## Larval Food Analysis and Qualitative Determination of Exoenzyme-Producing Gut Bacteria in Adult Ceratopogonid Midges (Diptera)

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### ABSTRACT

Biting midges are small nematoceros Diptera. *Culicoides* and *Dasyhelea* are two important genera of the family Ceratopogonidae. Larvae of *Culicoides innoxius* and *Dasyhelea aprojecta* are found in the semiaquatic moist habitat. The larvae feed on the small debris and habitat substrata. The materials consumed by these larvae aid in their development to become adult. The nutritional evaluation of the food material of larvae of *C. innoxius* and *D. aprojecta* was carried out to know the essential elements for their development. In the case of adult *Culicoides*, many species are hematophagous. However, the adult midges of the genus *Dasyhelea* are dependent on nectar and honeydew. Along with their digestive enzymes, exoenzyme-producing gut associated bacteria have also an important role in the digestion of these food materials. Digestion and metabolism of these food materials aid in insect maturation, immunity, reproduction, maintaining diapause, etc. Qualitative determination of the gut associated bacteria of adult *C. innoxius* and *D. flava* was accomplished to infer the role of bacteria supplementing the digestive enzymes.

**Keywords:** *Culicoides*, *Dasyhelea*, Larval food material, Proximate composition, Exoenzyme-producing gut bacteria, Qualitative determination.

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### INTRODUCTION

Family Ceratopogonidae is an enormously diverse group of small nematoceros midges, usually known as biting midges. In this family along with some other genera *Culicoides* Latreille [1], and *Dasyhelea* Kieffer [2], are important in having medico-veterinary and economic significance respectively. Many species of *Culicoides* are vectors of pathogenic protozoans, viruses, and filarial nematodes [3-5] causing Akabane, Blue Tongue disease, etc. [6, 7]. Some members of the genus *Dasyhelea* are pollinators of cocoa and rubber trees [8, 9]. In the case of female *Culicoides*, the adult stage is hematophagous, feeding on vertebrate blood, though there is report of natural sugar feeding habit also [10]. The larvae of some species of

*Culicoides* are trophic generalists [11]. Adult stages of both the sexes belonging to the genus *Dasyhelea* rely on nutrition from the honeydew and nectar [12], larval stages of this genus are primarily herbivorous actively feeding upon the plant and animal debris [13-15]. However, there is also evidence of carnivorous feeding habit of *D. pseudoincisurata* Waugh and Wirth, [16, 17]. Insects harbor a broad variety of microorganisms in their gut which help in numerous physiological functions [18]. There is also role of autochthonous bacteria in production of digestive enzymes for plant-derived polymers [19]. Digestion of lipid and protein may also be contributed by these microorganisms [20].

This article aims to evaluate the composition of food materials ingested by the larvae during

active feeding stage. So far, no investigation has been made for qualitative determination of autochthonous enzyme-producing gut bacteria of adults *Culicoides* and *Dasyhelea* which we have also done in this study.

## MATERIALS AND METHODS

### *Analysis of larval food (Figures 1a-1c)*

A site (Site: India, West Bengal, Nadia, Krishnanagar) was selected for the collection of food material. Rotting banana stems were found at the collection site and the presence of flying adults confirmed the breeding habitat. Two types (Sample 1 and Sample 2) of the rotting stems were cut out with the help of a fine knife, bagged in 45×40 cm clear plastic packets, and brought to the laboratory keeping the packets airtight. The samples were kept in a 30 cm × 25 cm × 4 cm plastic tray with a little amount of water just to make the sample moistened. A small amount of sample was taken and examined carefully for the presence of any larva. Some larvae were observed underneath the Olympus SZX 16 microscope (Japan) and found consuming their habitat substrata which are the upper thin membranous leaflets of the stem in both the samples. Few larvae were isolated and reared up to adult emergence. Upon the adult eclosion, the midges were identified under Carl Zeiss Stemi 2000 Stereozoom microscope (Germany) and confirmed their belonging to the genera *Culicoides* and *Dasyhelea*. Further species-level identification was confirmed after mounting in glass slides and observing under a Wild Leitz GMBH Trinocular microscope (Portugal). The upper thin membranous layer of both samples was isolated carefully by forceps, washed thoroughly in tap water, and sun dried for 3–4 days. The two dried samples (Sample 1 and Sample 2) were then put into a mixer grinder, and completely pulverized. The ground sample 1 and sample 2 were then stored in zipper plastic packets. Estimation of crude lipid, crude fiber, ash content (minerals), crude protein content, total free amino acid, and free fatty acid content of the food material of *Culicoides* and *Dasyhelea* larvae was done by proximate composition analysis. Proximate composition of the samples were analyzed following the standard methods of the Association of Official Analytical Chemists

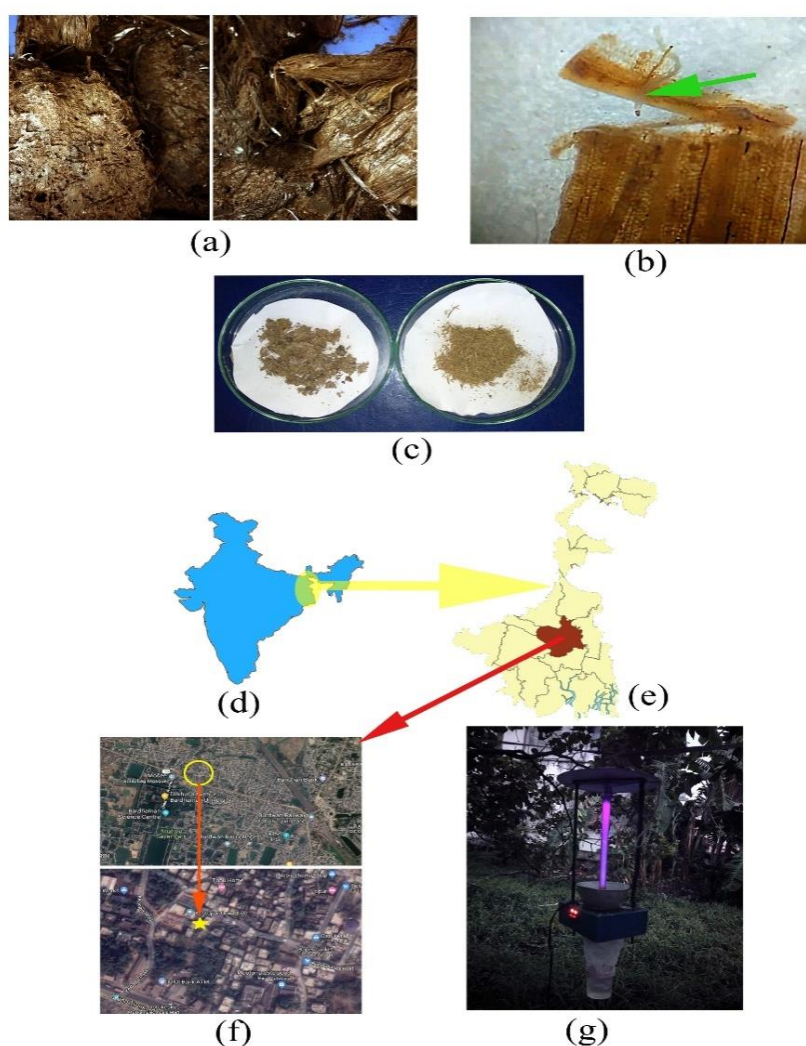
(AOAC)[21] – crude protein by micro Kjeldahl digestion and distillation, crude lipid by extracting the residue with 40-60°C petroleum ether in a Soxhlet apparatus (Pelican, Chennai, India), crude fiber as loss on ignition of dried lipid-free residue following digestion with 1.25% H<sub>2</sub>SO<sub>4</sub> and 1.25% NaOH and ash content by ignition at 550°C (6 hrs) in a Muffle furnace to constant weight. Total free amino acid and fatty acid contents were also measured [22, 23].

### *Qualitative determination of exoenzyme-producing ability of the gut-associated bacteria in adult midges (Figures 1d-1g)*

Battery operated UV light trap (8 W) was used for collecting the biting midges (Site: India, West Bengal, Purba Bardhaman, Burdwan, Keshtopur). The midges were collected in phosphate buffer saline. The collected insects were brought to the laboratory and sorted out quickly to avoid postmortem changes in physiology using a stereozoom microscope Carl Zeiss Stemi 2000 with the help of a fine “000” brush. The species were identified after mounting the genitalia on glass slides [24]. Midges of the genera *Culicoides* and *Dasyhelea* were separated and stored in plastic vials filled with phosphate buffer saline. For each species, the sample processing was performed separately. Samples of 0.05 g were weighed in Mettler Toledo Model - JS1203C/A71 (Switzerland) and washed thoroughly with distilled water. As a surface sterilizing agent 70% ethanol was used. The midges were beheaded beneath the stereozoom microscope using a fine needle and taken in an autoclaved plastic container of 2 ml and sterilized distilled volume of 20 times (i.e. 1 ml) of the weighed sample. IKA T10 basic ULTRA-TURRAX Model - T 10B (India) was used to homogenate the sample. The homogenized samples were stored in a refrigerator at 4°C in an autoclaved plastic container. The homogenates were serially diluted (1:10) with sterilized distilled water [25]. A total of three-step dilution 10<sup>-1</sup>–10<sup>-3</sup> was prepared for each species. In order to determine the culturable heterotrophic autochthonous aerobic/ facultative anaerobic microbial population, nutrient agar plates were prepared. The plates were stored in a refrigerator (4°C) until further study. A plate for each species having well-separated colonies with

distinguished growth is selected to prepare pure culture to get different bacterial isolates. After that, the colony characteristics of each isolate from pure culture were recorded and the colonies were transferred to new culture plates which were stored at 4°C in a refrigerator. Gram staining and endospore staining of bacteria were done. After that measurements of the bacteria were taken underneath compound light microscope (Wild Leitz GMBH, Portugal) using ocular and stage micrometer. For enzymatic analyses bacterial isolates were grown on

different culture media and stained: for amylase – cultured in starch agar then stained by Lugol's Iodine; for protease – cultured in agar with gelatin, peptone, and yeast extract then treatment by HCl + HgCl<sub>2</sub>; for cellulase – cultured in bacteriological agar with Carboxymethylcellulose (CMC), then stained by Congo Red; for lipase – cultured in Tributyrin agar and Tributyrin base. The plates were inspected for any halo zone, if present the zone characteristics were recorded.



**Figure 1.** a) Habitat substrata of Sample 1 (left) and 2 (right); b) Larva of biting midge in the habitat substratum; c) pulverized Sample 1 (left) and 2 (right); d) Map of India; e) Map of West Bengal showing the collection site district; f) Collection site of adult biting midges (Google Map: Imagery ©2022 Maxar Technologies, Imagery ©2022 CNES / Airbus, Maxar Technologies, Map data ©2022); g) collection of adult biting midges by UV light trap.

## RESULTS AND DISCUSSION

### Analyses of larval food

The larvae procured from the habitats were identified as *Culicoides (Hoffmania) innoxius* Sen

and Das Gupta, [26] and *Dasyhelea (Pseudoculicoides) aprojecta* Brahma, Chatterjee and Hazra, [27] upon adult emergence. The result of larval food content (habitat substrata) analysis is depicted in **Table 1**.

**Table 1.** Proximate composition of Sample 1 and Sample 2 (g/100g) consumed by the *Culicoides innoxius* and *Dasyhelea aprojectalarvae* respectively. Data expressed as mean  $\pm$  standard error (n = 3).

Sl. No.	Nutrients	Sample 1	Sample 2
1.	Crude Lipid	5.24 $\pm$ 0.14	9 $\pm$ 0.02
2.	Crude Protein	13.14 $\pm$ 0.35	11.74 $\pm$ 0.32
3.	Crude Fiber	35.96 $\pm$ 0.92	37.63 $\pm$ 1.01
4.	Ash content (minerals)	4.01 $\pm$ 0.11	0.79 $\pm$ 0.01
5.	Nitrogen Free Extract (NFE)	41.63 $\pm$ 1.02	40.82 $\pm$ 1.3
6.	Free amino acids (FAA)	0.88 $\pm$ 0.01	0.71 $\pm$ 0.01
7.	Free fatty acids (FFA)	0.77 $\pm$ 0.01	0.61 $\pm$ 0.001

#### Qualitative determination of exoenzyme-producing ability of the gut associated bacteria in adult midges (Figure 2)

The species used for enzymatic determination are identified as females of *Culicoides (Hoffmania) innoxius* Sen and Das Gupta, [26] and females of *Dasyhelea (Prokempia) flava* Carter, Ingram and Macfie, [28].

In the case of both species colonies of the concentration  $10^{-1}$  were overlapped and uncountable. Colonies of concentration  $10^{-2}$  were separated with distinct margins and variations, so they were selected for preparation of pure culture (In both *Culicoides innoxius* and *Dasyhelea flava*  $10^{-2}$  concentration produces eight morphologically distinct colonies) (Table 2). Least numbers of colonies were observed in concentration  $10^{-3}$ .

Only six isolates of *Culicoides innoxius* and *Dasyhelea flava* each (Cu<sub>1-3</sub>, Cu<sub>5-7</sub>; Da<sub>1-3</sub>, Da<sub>5-7</sub>) were taken for downstream analysis for better growth in pure culture. Among the six isolates retrieved from both the species (Cu<sub>1-3</sub>, Cu<sub>5-7</sub>; Da<sub>1-3</sub>, Da<sub>5-7</sub>) two isolates (Cu<sub>3</sub>, Cu<sub>7</sub>) were found as Gram-positive while the other four (Cu<sub>1</sub>, Cu<sub>2</sub>, Cu<sub>5-6</sub>) were found to be Gram-negative in the case of *Culicoides innoxius*, and three isolates (Da<sub>1-3</sub>) were found as Gram-positive while other three (Da<sub>5-7</sub>) were found as Gram-negative in the case of *Dasyhelea flava*. All the isolates belonging to *Culicoides* are bacilli and isolates belonging to *Dasyhelea* consist of a mixture of cocci form (Da<sub>1</sub>) and bacilli (Isolates Da<sub>2-3</sub>, Da<sub>5-7</sub>). The length and width of bacilli retrieved from *Culicoides* range from 1.4–2.76  $\mu$ m and 0.60–0.74  $\mu$ m respectively (Table 3); the length and width of bacilli retrieved from *Dasyhelea* range from 1.82–2.8  $\mu$ m and 0.69–0.74  $\mu$ m

respectively and the diameter of cocci form ranges between 0.62 and 0.64  $\mu$ m (Table 3). All six isolates from both species were found as non-endospore-forming bacteria.

#### Determination of enzyme-producing bacteria in adult *Culicoides innoxius* (Figures 2a–2d; Tables 4 and 5)

**Amylase.** Halo zones were observed at the periphery of colonies Cu<sub>1-7</sub>. So, six isolates were found as amylase positive.

**Protease.** Halo zones were observed at the periphery of colonies Cu<sub>1-3</sub> and Cu<sub>5-7</sub>. So, six isolates were found as protease positive.

**Cellulase.** Halo zones were observed at the periphery of colonies Cu<sub>1-2</sub>. So, two isolates were found cellulase positive.

**Lipase.** Halo zones were observed at the periphery of colonies Cu<sub>1-3</sub> and Cu<sub>5-7</sub>. So, six isolates were found as lipase positive.

#### Determination of enzyme-producing bacteria in adult *Dasyhelea flava* (Figures 2e–2h; Tables 4 and 5)

**Amylase.** Halo zones were observed at the periphery of colonies Da<sub>1-3</sub> and Da<sub>6-7</sub>. So, four isolates were found as amylase positive.

**Protease.** Halo zones were observed at the periphery of colonies Da<sub>1</sub> and Da<sub>6-7</sub>. So, three isolates were found as protease positive.

**Cellulase.** No halo zone was observed around the periphery of any colony. Hence, all six isolates (Da<sub>1-3</sub> and Da<sub>5-7</sub>) were found cellulase negative.

**Lipase.** Halo zones were observed at the periphery of all colonies (Da<sub>1-3</sub> and Da<sub>5-7</sub>) So, six isolates were found as lipase positive.

In past, nutritional and chemical properties of the larval habitats of certain species of *Culicoides* were evaluated, and categorization of the species of *Culicoides* was done based on inhabiting high nutrients, moderate to high nutrients, wide nutrient range, moderate to low nutrient, and low nutrient zone [29]. Larvae of *Dasyhelea* feed on algae and fungi [13]. But carnivorous feeding habit of *D. ampullariae* Macfie, [30] was also reported [31] which consumes mosquito larvae. There was no record regarding the nutrient contents in the habitat substrata which are consumed by the larvae of *Dasyhelea* before this study. The larval stages of *Culicoides* and *Dasyhelea* actively feed upon the debris, organic matter, and substrata present in their habitat. In this study, proximate composition analysis reveals the number of

different nutrients in the food content. High crude fiber quantity has been found in both samples. Moderate quantities of crude protein and crude lipid are also observed which may be crucial for larval maturation. High value of Nitrogen Free Extract (NFE) corresponds to a high amount of carbohydrate which may be important for larval growth. Both types of

habitat substrate samples reflect low Free Amino Acids (FAA) and low Free Fatty Acids (FFA) quantity referring to their little requirement for larval development. Finally, a low to moderate amount of minerals (ash content) may be sufficient for sustaining metabolic activity in the larval stages of the *Culicoides innoxius* and *Dasyhelea aprojecta*.

**Table 2.** The colony characteristics of eight bacterial isolates from *Culicoides innoxius* and *Dasyhelea flava*.

Characteristics	Isolate 1		Isolate 2		Isolate 3		Isolate 4		Isolate 5		Isolate 6		Isolate 7		Isolate 8	
	(Cu <sub>1</sub> )	(Da <sub>1</sub> )	(Cu <sub>2</sub> )	(Da <sub>2</sub> )	(Cu <sub>3</sub> )	(Da <sub>3</sub> )	(Cu <sub>4</sub> )	(Da <sub>4</sub> )	(Cu <sub>5</sub> )	(Da <sub>5</sub> )	(Cu <sub>6</sub> )	(Da <sub>6</sub> )	(Cu <sub>7</sub> )	(Da <sub>7</sub> )	(Cu <sub>8</sub> )	(Da <sub>8</sub> )
<b>Form</b>	Circular	Circular	Circular	Circular	Circular	Irregular	Circular	Circular	Irregular	Acute circular	Acute circular	Acute circular	Circular	Circular	Irregular	Irregular
<b>Size</b>	Punctiform	Punctiform	14-15mm	10-15mm	10-12mm	10-15mm	10-15mm	20-27mm	Punctiform	Punctiform	12-15mm	14-15mm	20-24mm	28-30mm	10-14mm	10-11mm
<b>Elevation</b>	Flat	Raised	Flat	Flat	Flat	Flat	Flat	Flat	Raised	Raised	Raised	Raised	Raised	Raised	Flat	Flat
<b>Margin/Border</b>	Undulate	Entire	Entire	Entire	Undulate	Undulate	Irregular	Irregular	Entire	Entire	Entire	Entire	Entire	Entire	Irregular	Irregular
<b>Surface</b>	Rough	Smooth	Smooth	Smooth	Rough	Rough	Dull	Dull	Glistening	Glistening	Glistening	Glistening	Glistening	Rough	Rough	Rough
<b>Opacity</b>	Opaque	Opaque	Opaque	Opaque	Semitranslucent	Semitranslucent	Translucent	Translucent	Opaque	Opaque	Opaque	Opaque	Opaque	Opaque	Semitranslucent	Semitranslucent
<b>Color</b>	Yellowish white	Yellow	White	White	White	White	Dull white	Dull white	Yellow	White	Yellow	Bright yellow	Yellowish white	Yellowish white	Dull white	Grey

**Table 3.** Characteristics of bacteria from pure cultures and their endospore-forming ability.

Isolates	Gram Positive/ Negative	Bacilli/ Cocci	Length/ Diameter (µm)	Width (µm)	Endospore
<b>Cu<sub>1</sub></b>	Negative	Bacilli	1.42	0.70	Absent
<b>Cu<sub>2</sub></b>	Negative	Bacilli	1.80	0.64	Absent
<b>Cu<sub>3</sub></b>	Positive	Bacilli	1.80	0.62	Absent
<b>Cu<sub>5</sub></b>	Negative	Bacilli	1.84	0.60	Absent
<b>Cu<sub>6</sub></b>	Negative	Bacilli	2.74	0.74	Absent
<b>Cu<sub>7</sub></b>	Positive	Bacilli	1.76	0.72	Absent
<b>Da<sub>1</sub></b>	Negative	Cocci	0.64	-	Absent
<b>Da<sub>2</sub></b>	Positive	Bacilli	1.84	0.73	Absent

<b>Da<sub>3</sub></b>	Negative	Bacilli	2.76	0.69	Absent
<b>Da<sub>5</sub></b>	Negative	Bacilli	2.80	0.70	Absent
<b>Da<sub>6</sub></b>	Positive	Bacilli	1.84	0.74	Absent
<b>Da<sub>7</sub></b>	Positive	Bacilli	1.82	0.73	Absent

**Table 4.** Extracellular enzyme producing ability of different isolates (“+” = positive, “-” = negative).

	<b>Amylase</b>	<b>Protease</b>	<b>Cellulase</b>	<b>Lipase</b>
<b>Cu<sub>1</sub> / Da<sub>1</sub></b>	+ / +	+ / +	+ / -	+ / +
<b>Cu<sub>2</sub> / Da<sub>2</sub></b>	+ / -	+ / -	+ / -	+ / +
<b>Cu<sub>3</sub> / Da<sub>3</sub></b>	+ / +	+ / -	- / -	+ / +
<b>Cu<sub>5</sub> / Da<sub>5</sub></b>	+ / -	+ / -	- / -	+ / +
<b>Cu<sub>6</sub> / Da<sub>6</sub></b>	+ / +	+ / +	- / -	+ / +
<b>Cu<sub>7</sub> / Da<sub>7</sub></b>	+ / +	+ / +	- / -	+ / +

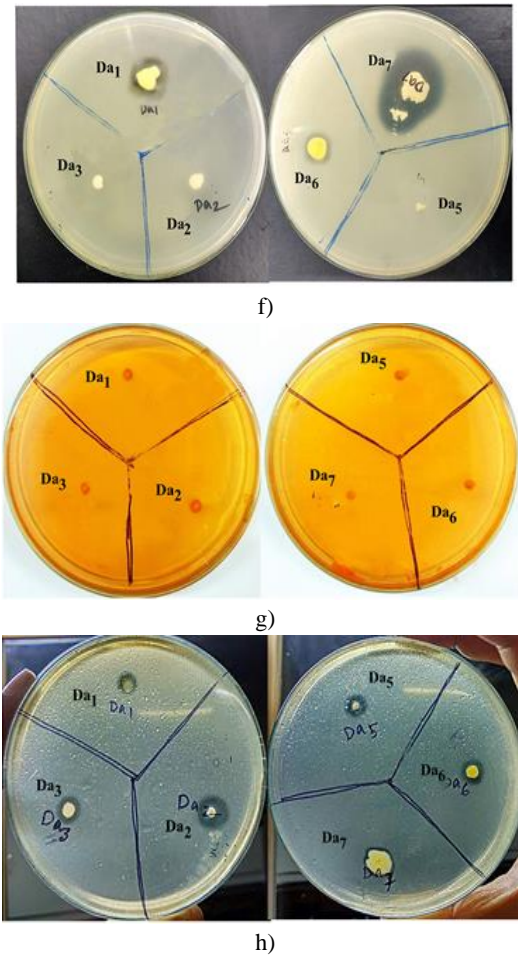
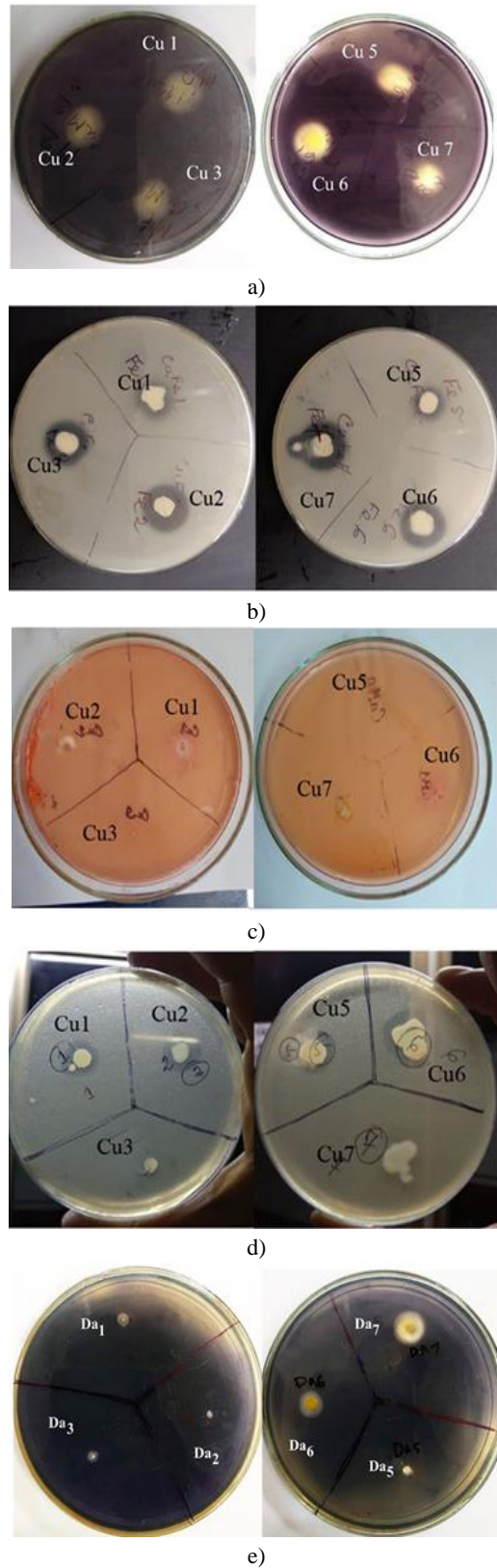
**Table 5.** Halo zone characteristics (Diameter: 1-4mm = +, 5-8mm = ++, 9-12mm = +++, 13-16mm = +++++, 17-20mm = ++++++ and 21-24mm = +++++++ ; margin: very diffused = VD, diffused = D, prominent = P, - = negative).

<b>Enzymes</b>	<b>Isolates</b>											
	<b>Cu<sub>1</sub></b>	<b>Da<sub>1</sub></b>	<b>Cu<sub>2</sub></b>	<b>Da<sub>2</sub></b>	<b>Cu<sub>3</sub></b>	<b>Da<sub>3</sub></b>	<b>Cu<sub>5</sub></b>	<b>Da<sub>5</sub></b>	<b>Cu<sub>6</sub></b>	<b>Da<sub>6</sub></b>	<b>Cu<sub>7</sub></b>	<b>Da<sub>7</sub></b>
<b>Amylase</b>	+++ , D	++ , VD	+++ , P	-	+++ , D	+ , D	+++ , P	-	+++ , P	++ , D	+++ , P	+++ , P
<b>Protease</b>	+++ , P	+++ , D	+++ , P	-	+++ , P	-	+++ , P	-	+++ , P	+++ , D	+++ , P	++++++ , P
<b>Cellulase</b>	++ , D	-	+ , D	-	-	-	-	-	-	-	-	-
<b>Lipase</b>	++ , P	++ , P	++ , P	+++ , P	++ , D	++ , P	+++ , P	++ , P	+++ , P	++ , P	+++ , D	+++ , P

*Culicoides innoxius* has a potential role in spreading BTV through its blood-feeding habit in cattle [6, 7]. Adult *Dasyhelea* feeds on honeydew and sugar solutions of plants [16]. Adult biting midges may also visit flowers for nectar. Nevertheless, no information was available regarding extracellular enzyme-producing bacteria in the gut of adult *Culicoides* and *Dasyhelea*. Starch, a unique source of energy for these midges can provide the vigor for sustained flight and reproduction. In this study, six and four bacterial isolates were recorded as amylase positive in females of *Culicoides innoxius* and *Dasyhelea flava* respectively, indicating the positive role of the gut bacterial colonies in supplementing the enzyme. A high amount of the phloem sap proteins may have a role in stress and defense reactions in the phloem feeder insects [32], which also corroborates with our present study as we got three protease positive bacterial isolates from *Dasyhelea flava*. However, six protease-positive isolates indicate a protein-rich diet of *Culicoides innoxius* inferring their possible hemtrophagous habit upon the vertebrate host. Cellulose is derived from D-glucose units and condenses through  $\beta$ -1, 4-glycosidic linkage. As cellulose is

the structural part of a plant cell wall, insects that feed on fluids of xylem or phloem cannot get access to the cellulose content. In the case of *Culicoides innoxius*, only two isolates were found as cellulase positive which leads us to assume its little dependency on cellulose rich food. However, in case of *Dasyhelea flava* no cellulase-producing bacteria was detected confirming that they feed on nectar, honeydew, etc. [12] which lack cellulose content. So, the feeding habit of adult *Dasyhelea flava* might have made their gut unfavorable for the growth of cellulase-producing bacteria. Triglycerols stored in adipose tissue are a major form of energy storage in animals. Insect adipocytes can store a large amount of lipid reserves as cytoplasmic lipid droplets [33]. Insect embryogenesis is highly dependent upon lipid metabolism and during their development, insect oocytes increase their lipid content several fold in a very short period [33]. Insect growth and reproduction, synthesis of trehalose and proline, and insect immune response rely upon lipid metabolism and lipids also provide the energy needed during diapause [34]. Fatty acids stored as triglycerides in anhydrous form can be used for energy production through  $\beta$ -oxidation [35].

The present study validates the vast role of lipid utilization by the *Culicoides innoxius* and *Dasyhelea flava* midges as all six isolates retrieved from both species were detected as lipase positive. Thus, gut bacteria aid in the digestion of lipid in the adult actively.



**Figure 2.** Qualitative Determination of exo-enzyme producing autochthonous gut associated bacteria in adult *Culicoides innoxius* Sen and Das Gupta and *Dasyhelea flava* Carter, Ingram and Macfie: a) amylase; b) protease;c) cellulase and d) lipase producing bacterial determination of *Culicoides innoxius*; e) amylase; f) protease; g) cellulase and h) lipase producing bacterial determination of *Dasyhelea flava*.

**CONCLUSION**

The larvae of *Culicoides* are reported as trophic generalists and larvae of *Dasyhelea* are stated to usually feed on algae and fungi but the present investigation with the larvae of *Culicoides (Hoffmania) innoxius* [26] and *Dasyhelea (Pseudoculicoides) aprojecta* [27] are found to feed on plant derived organic habitat substrata. So, an attempt was made to get an idea about the overall nutrient content of the probable larval food material by analyzing the proximate composition of the substances. The result reveals the presence of a high quantity of crude fiber, moderate quantities of crude protein, crude lipid, low to moderate amount of minerals, and a high amount of carbohydrate

which may be vital for larval growth and metabolic activity. The result also refers to fewer requirements of Free Amino Acids (FAA) and Free Fatty Acids (FFA) as the value of these is very low.

Qualitative determination of exoenzyme-producing ability of the gut associated bacteria in adult *Culicoides innoxius* and *Dasyhelea flava* [28] midges indicate the presence of amylase, protease, cellulase, and lipase-producing bacteria in former; however, only cellulase producing bacteria are wanting in latter. Among the six bacterial isolates grown from the gut of *C. innoxius* all are positive in producing amylase, protease, and lipase, and only two isolates produce cellulase. However, in *D. flava*, all six, four and three isolates have been recorded as lipase, amylase, and protease positive respectively and no cellulase producing gut bacteria has been detected, reason may be due to their plant sap and nectar feeding habit. These gut bacteria have their role in supplementing the enzymes for digestion.

Detailed analysis of the larval food materials and association of symbiotic gut bacteria of adult biting midges will help the future workers to rear the insects in large number which will eventually help to decipher the biology of medico-veterinary important *Culicoides* spp. and economically important *Dasyhelea* spp. However, rearing of economically important *Dasyhelea* spp. will have a havoc impact on cocoa and rubber production.

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## REFERENCES

1. Latreille PA. Genera crustaceorum et insectorum secundum ordinem naturalem in familias disposita, iconibus exemplisque plurimis explicata. Vol. 4. Paris and Strasbourg, France; 1809, pp. 399.
2. Kieffer JJ. Nouvelles descriptions de chironomides obtenus d'éclosion. Bull Soc Hist nat Metz. 1911;27:1-60.
3. Mellor PS, Boorman J, Baylis M. *Culicoides* biting midges: their role as arbovirus vectors. Annu Rev Entomol. 2000;45(1):307-40. doi:10.1146/annurev.ento.45.1.307
4. Borkent A. "Ceratopogonidae", in Marquart WC. (Editor), Biology of disease vectors. 2nd ed. Elsevier Press, Amsterdam; 2005.785 p.
5. Prasad G, Sreenivasulu D, Singh KO, Mertens PPC, Maan S. Bluetongue in the Indian subcontinent. In: Mellor PS, Baylis M, Mertens PPC. (Editors), Bluetongue. Elsevier - Academic Press, London; 2009. pp. 167-95.
6. Kettle DS. Biology and bionomics of bloodsucking ceratopogonids. Annu Rev Entomol. 1977;22:33-51. doi:10.1146/annurev.en.22.010177.000341
7. Linley JR, Hoch AL, Pinheiro FP. Biting midges (Diptera: Ceratopogonidae). J Med Entomol. 1983;20(4):347-64. doi:10.1093/jmedent/20.4.347
8. Lee KM, Wirth WW, Chan KL. A new species of *Dasyhelea* midge reared from drains in Singapore (Diptera: Ceratopogonidae). Proc Entomol Soc Wash. 1989;91(3):452-7.
9. Wirth WW, Waugh WT. Five new Neotropical *Dasyhelea* midges (Diptera: Ceratopogonidae) associated with culture of cocoa. Stud Entomol. 1976;19:223-36.
10. Kaufman C, Mathis A, Vorburger C. Sugar-feeding behaviour and longevity of



- European *Culicoides* biting midges. Med Vet Entomol. 2015;29(1):17-25. doi:10.1111/mve.12086
11. Aussel JP, Linley JR. Natural food and feeding behavior of *Culicoides furens* larvae (Diptera: Ceratopogonidae). J Med Entomol. 1994;31(1):99-104. doi:10.1093/jmedent/31.1.99
  12. Dominiak P. Biting midges of the genus *Dasyhelea* Kieffer (Diptera: Ceratopogonidae) in Poland. Pol J Entomol. 2012;81(3):211-304. doi:10.2478/v10200-012-0009-8
  13. Mullen GR, Hribar LJ. Biology and feeding behavior of Ceratopogonid larvae (Diptera: Ceratopogonidae) in North America. Bull Soc Vector Ecol. 1988;13(1):60-81.
  14. Lieven AF. Functional morphology and phylogeny of the larval feeding apparatus in the Dasyheleinae and Forcipomyiinae (Diptera, Ceratopogonidae). Deut Entomol Z. 1998;45(1):49-64. doi:10.1002/mmnd.19980450107
  15. Ronderos MM, Spinelli G, Huerta H, Díaz F. Immature stages of two Neotropical species of *Dasyhelea* Kieffer, 1911 (Diptera: Ceratopogonidae). Trans Am Entomol Soc. 2003;129(2):295-330.
  16. Waugh WT, Wirth WW. A revision of the genus *Dasyhelea* Kieffer of the eastern United States north of Florida (Diptera: Ceratopogonidae). Ann Entomol Soc Am. 1976;69(2):219-47. doi:10.1093/AESA/69.2.219
  17. Hribar LJ. Biological and morphological notes on *Dasyhelea pseudoincisurata* (Diptera: Ceratopogonidae). Entomol News. 1998;109(4):282-4.
  18. Dillon RJ, Dillon VM. The gut bacteria of insects: non-pathogenic interactions. Ann Rev Entomol. 2004;49(1):71-92. doi:10.1146/annurev.ento.49.061802.123416
  19. Douglas AE. The microbial dimension in insect nutritional ecology. Funct Ecol. 2009;23(1):38-47. doi:10.1111/j.1365-2435.2008.01442.x
  20. Visotto LE, Oliveira MG, Guedes RNC, Ribon AOB, Good-God PIV. Contribution of gut bacteria to digestion and development of the velvet bean caterpillar, *Anticarsia gemmatalis*. J Insect Physiol. 2009;55:185-191. doi:10.1016/j.jinsphys.2008.10.017
  21. AOAC. Official methods of analysis. Arlington VA. (16th Ed), Association of Official Analytical Chemist, New York;1990. 1134 p.
  22. Moore S, Stein WH. Photometric ninhydrin method for use in the chromatography of amino acids. J Biol Chem. 1948;176:367-88. doi:10.1016/S0021-9258(18)51034-6
  23. Cox HE, Pearson D. The Chemical analysis of foods. Chemical Publishing Co, New York, United States of America; 1962. 3-10 pp.
  24. Wirth WW, Marston N. A method for mounting small insects on microscopic slides in Canada balsam. Ann Entomol Soc Am. 1968;61(3):783-4. doi:10.1093/aesa/61.3.783
  25. Beveridge MCM, Sikdar PK, Frerichs GN, Millar S. The ingestion of bacteria in suspension by the common carp *Cyprinus carpio* L. J Fish Biol. 1991;39(6):825-31. doi:10.1111/j.1095-8649.1991.tb04412.x
  26. Sen P, Das Gupta SK. Studies on Indian *Culicoides* (Ceratopogonidae: Diptera). Ann Entomol Soc Am. 1959;52(5):617-30.
  27. Brahma S, Chatterjee S, Hazra N. Three new species of *Dasyhelea* Kieffer and new record of *D. flaviformis* Carter, Ingram and Macfie (Diptera: Ceratopogonidae) from the Deltaic Proper of Gangetic West Bengal, India. J Insect Biodivers. 2020;016(2):055-80. doi:10.12976/JIB/2020.16.2.3
  28. Carter HF, Ingram A, Macfie JWS. Observations on the ceratopogonine midges of the Gold Coast with descriptions of new species Part IV. Ann Trop Med Parasitol. 1921;15:177-212. doi:10.1080/00034983.1921.11684266
  29. Battle FV, Turner Jr EC. Some nutritional and chemical properties of the larval habitats of certain species of *Culicoides* (Diptera: Ceratopogonidae). J Med Entomol. 1972;9(1):32-5. doi:10.1093/jmedent/9.1.32
  30. Macfie JWS. Report on a collection of Ceratopogonidae from Malaya. Annals of Tropical Medicine and Parasitology. 1934;28(2):177-94. doi:10.1080/00034983.1934.11684809
  31. Meng LK, Lok CK. The biology of *Dasyhelea ampullariae* in monkey cups at Kent Ridge

- (Diptera: Ceratopogonidae). *J Singapore Nad Acad Sci.* 1985;14:6-14.
32. Kehr J. Phloem sap proteins: their identities and potential roles in the interaction between plants and phloem-feeding insects. *J Exp Bot.* 2006;57(4):767-74. doi:10.1093/jxb/erj087
33. Arrese EL, Soulages JL. Insect fat body: Energy, metabolism, and regulation. *Ann Rev Entomol.* 2010;55:207-25. doi:10.1146/annurev-ento-112408-085356
34. Hahn DA, Denlinger DL. Meeting the energetic demands of insect diapause: nutrient storage and utilization. *J Insect Physiol.* 2007;53(8):760-73. doi:10.1016/j.jinsphys.2007.03.018
35. Athenstaedt K, Daum G. The life cycle of neutral lipids: synthesis, storage and degradation. *Cell Mol Life Sci.* 2006;63(12):1355-69. doi:10.1007/s00018-006-6016-8