

## Population Trend of Onion Thrips and Its Botanical Approach to Sustainable Management

Ashish Kumar<sup>1</sup>, Ankush Raut<sup>1\*</sup>, Paritosh Tripathi<sup>2</sup>, Najitha Banu<sup>3</sup>

<sup>1</sup>Department of Entomology, School of Agriculture, Lovely Professional University, Phagwara, Punjab, India.

<sup>2</sup>Department of Entomology, Chandra Shekhar Azad University of Agriculture and Technology, Kanpur, Uttar Pradesh, India.

<sup>3</sup>Department of Zoology, School of Bioengineering and Bioscience, Lovely Professional University, Phagwara, Punjab, India.

### ABSTRACT

This research is about onion (*Allium cepa* L.) which is an important commonly grown vegetable in kitchen gardens and for commercial purposes throughout the globe its crucial. Plant phytochemicals are playing an important role in pest management, and when combined with synthetic pesticides, they increase toxicity due to a synergistic effect. The current study compares effectiveness of *Lantana camera* leaf extract and its combination with diamethoate 30EC against onion thrips were compared. It has resulted that thrips colonization began on the 9th SMW and peaked on the 15th SMW. Thrips colonization was maximum at high temperature and low rainfall but relative humidity depleting the thrip population. All treatments were found to be significantly more effective than the control against thrips. *Lantana camera* leaves extracts at 50 g L<sup>-1</sup> as the sole and in combination with dimethoate 30EC between 150 and 250 mL ha<sup>-1</sup> were found to be a promising combination.

**Keywords:** *Thrips tabaci*, *Lantana camera*, Sustainability, Management, Population dynamics.

**HOW TO CITE THIS ARTICLE:** Kumar A, Raut A, Tripathi P, Banu N. Population Trend of Onion Thrips and Its Botanical Approach to Sustainable Management. Entomol Appl Sci Lett. 2022;9(3):25-31. <https://doi.org/10.51847/LlgFcSiV6X>

**Corresponding author:** Ankush Raut

**E-mail** ✉ [ankushento@gmail.com](mailto:ankushento@gmail.com); [ankush.19876@lpu.co.in](mailto:ankush.19876@lpu.co.in)

**Received:** 26/03/2022

**Accepted:** 10/08/2022

### INTRODUCTION

Onion is a valuable cash crop grown primarily by smallholder farmers under irrigation, with the total area covered increasing over time [1, 2]. A poor cropping system, a scarcity of resistant varieties, disease and insect pests, and other constraints have all contributed to lower bulb production [3]. *Thrips tabaci* was one of the most common pest [4], causing 10-60% economic loss in onions depending on various factors [5] and up to 100% in some crops [6, 7]. *T. tabaci* is a cosmopolitan and polyphagous pest with nymphs and adults found between leaf sheaths that attack over 391 plant species belongs to different 64 families [8]. They both extensively scrape the leaves, sucking the cell

sap and leaving silver patches and streaks that eventually dry the leaves [9, 10]. To compensate for this loss, farmers depend entirely on synthetic pesticide formulations, but their indiscriminate use has an impact on crop costs, and human and environmental health [11, 12]. Furthermore, sucking pests such as thrips rapidly developed insecticide resistance and became ineffective [13]. As a result, botanical derivatives offer an alternative option to overcome the resistance problem.

Plants and plant products have been used in agriculture in China, Egypt, Greece, and India since ancient times [14]. Furthermore, plant extracts are becoming more popular as organic products and could be used as an alternative pest management strategy [15]. Many plant extracts have been investigated for their toxic,

antifeedant, ovicidal, and repellent properties [16-20]. *Lantana camera* is a tropical ornamental weed that grows as an erect shrub. Due to the presence of secondary metabolites [21, 22], it has the potential to manage stored grain pests [23], lepidopteran pests [24], and sucking pests [25, 26]. There is very little information available on *L. camera* extract against onion thrips, so the current study focused on learning about thrips population buildup on onions and evaluating *L. camera* leaf extract against onion thrips.

## MATERIALS AND METHODS

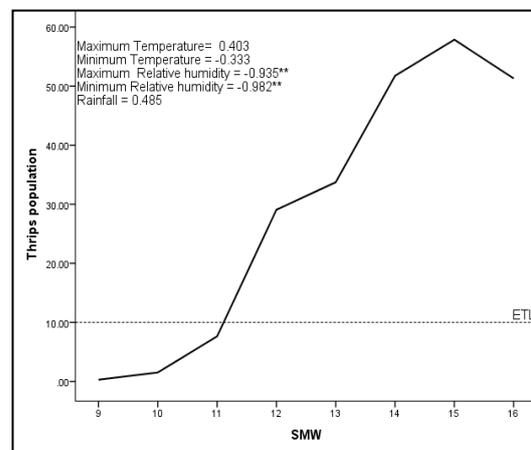
A separate field experiment was conducted to better understand the onion thrips trend and its management during the rabi season. A three-leaf stage of Punjab Naroya onion seedling was transplanted in 15 x 15 cm spacing in 216 m<sup>2</sup> areas. A transplanted area was kept completely chemical free to allow natural population build-up. To evaluate *L. camera* leaf extract, onion seedlings were transplanted in a 3x3 m<sup>2</sup> area. This experiment used a Randomized Block Design (RBD), with seven treatments, including a control, replicated three times. The treatments were *Lantana camera* leaf extract @ 50 g L<sup>-1</sup> (lower dose), 100 g L<sup>-1</sup> (standard dose), and 150 g L<sup>-1</sup> (higher dose). Recommended pesticides like Dimethoate 30% EC @ 250 mL ha<sup>-1</sup> were used in combination with each dose of leaf extract to compare with the absolute check. A recommended package of practices was adapted for uniform growth of the onion crop. Both nymph and adult of the thrips population were counted after every three-day interval from randomly selected 10 plants with the help of a hand lens.

Thrips count data was normalized using square root transformation, and the same data was used in various statistical analyses based on the objectives. An analysis of variance (ANOVA) based on Turkey's test was used to compare treatments. A correlation model developed between thrips populations and environmental factors. All the statistical analysis was performed in SPSS (version 22).

## RESULTS AND DISCUSSION

### Population dynamics and its relation with environmental factors

The population of *Thrips tabaci* appeared in the 9th standard meteorological week (SMW) and remained until crop maturity (16th SMW). The average population per five plants ranged from 0.28 to 57.87 thrips. The thrips population grew exponentially, reaching a peak at the 15th SMW (57.87 thrips/five plants) and then gradually declining with crop maturity (**Figure 1**). Temperature and rainfall were clearly shown to positively influence the population of *T. tabaci* ( $r=0.403$  and  $r=0.485$ ) respectively, whereas humidity ( $p=0.001$ ) significantly inhibited the population (**Figure 1**).



**Figure 1.** Population dynamics of *Thrips tabaci* and its relation with abiotic factors. (Dotted line on Y axis indicate economic threshold level)

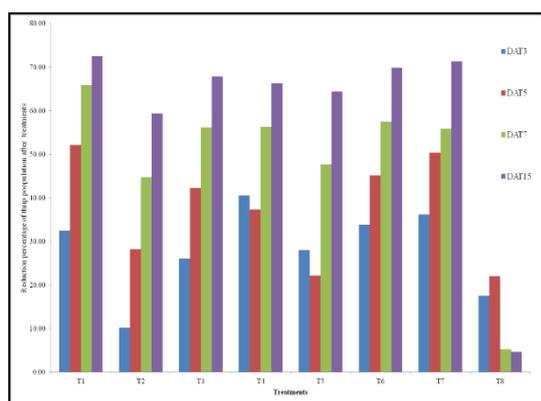
### Bio-efficacy of *lantana camera* and its combination with Diamethoate

The economic threshold level was set as 10 thrips per plant and pesticides were applied when the thrips population was well colonized on crops (**Figure 2**). All the treatments were performed significantly better at three days after treatments (DAT) (F 7, 2 =11.73,  $P<0.05$ ), 5 DAT (F 7, 2 =19.06,  $P<0.05$ ), 7DAT (F 7, 2 =40.54,  $P<0.05$ ) and 15 DAT (F 7, 2 =54.03,  $P<0.05$ ) than control (**Table 1**). *Lantana camera* leaf extract @ 50 g L<sup>-1</sup> reduced 32.46% thrips population at 3DAT, 5DAT (52.10 percent), 7DAT (65.93 percent), and 15 DAT (72.55 percent) when compared to higher doses (**Figure 2**). However, the *L. camera* leaf extract @ 50 g L<sup>-1</sup> in combination with Dimethoate 30% EC @ 250 mL ha<sup>-1</sup> was economically minimizing 40.51%, 37.35%, 56.32% and 66.21% of thrips population at 3DAT, 5DAT, 7DAT and 15DAT

respectively (**Figure 2**). However, among the all treatments, the sole application of Diamethoate 30% EC @ 250 mL ha<sup>-1</sup> was found highly superior to the other treatments against onion thrips.

**Table 1.** Effect of *Lantenna camera* leaf extract and its combination with Dimethoate spray against thrips population

	Post-treatment thrip population (number)				
	DAT3	DAT5	DAT7	DAT15	
T <sub>1</sub>	49.9	33.7 <sup>b</sup>	23.9 <sup>bc</sup>	17.0 <sup>b</sup>	13.7 <sup>b</sup>
T <sub>2</sub>	44.0	39.5 <sup>b</sup>	31.6 <sup>b</sup>	24.3 <sup>b</sup>	17.9 <sup>b</sup>
T <sub>3</sub>	51.3	37.9 <sup>b</sup>	29.6 <sup>bc</sup>	22.5 <sup>b</sup>	16.5 <sup>b</sup>
T <sub>4</sub>	50.6	30.1 <sup>b</sup>	31.7 <sup>b</sup>	22.1 <sup>b</sup>	17.1 <sup>b</sup>
T <sub>5</sub>	40.7	29.3 <sup>b</sup>	31.7 <sup>b</sup>	21.3 <sup>b</sup>	14.5 <sup>b</sup>
T <sub>6</sub>	44.7	29.6 <sup>b</sup>	24.5 <sup>bc</sup>	19.0 <sup>b</sup>	13.5 <sup>b</sup>
T <sub>7</sub>	41.5	26.5 <sup>b</sup>	20.6 <sup>c</sup>	18.3 <sup>b</sup>	11.9 <sup>b</sup>
T <sub>8</sub>	48.9	59.3 <sup>a</sup>	62.7 <sup>a</sup>	51.6 <sup>a</sup>	51.3 <sup>a</sup>
SEm (±)	3.216	3.377	2.863	1.761	1.765
P value	NS	10.243*	8.684*	5.340*	5.353*



**Figure 2.** Reduction percentage of thrips population after treatment (T<sub>1</sub>= *Lantana camera* leaf extract @ 50 g L<sup>-1</sup>, T<sub>2</sub>= *Lantana camera* leaf extract @ 100 g L<sup>-1</sup>, T<sub>3</sub>= *Lantana camera* leaf extract @ 150 g L<sup>-1</sup>, T<sub>4</sub>= *Lantana camera* leaf extract @ 50 g L<sup>-1</sup>+ Dimethoate 30% EC @ 250 ml/ha, T<sub>5</sub>= *Lantana camera* leaf extract @ 100 g L<sup>-1</sup>+ Dimethoate 30% EC @ 250 mL ha<sup>-1</sup>, T<sub>6</sub>= *Lantana camera* leaf extract @ 150 g L<sup>-1</sup>+ Dimethoate 30% EC @ 250 mL ha<sup>-1</sup>, T<sub>7</sub>= Dimethoate 30% EC @ 250 mL ha<sup>-1</sup>, T<sub>8</sub>= Control). DAT= days after treatments, <sup>a</sup>Different letter(s) showed significant differences between treatments (Turkey's  $p \leq 0.05$ ).

Onion thrips, *Thrips tabaci* is a serious and regular pest of onion distributed throughout the world. The immature stage of thrips causes direct damage by lacerating the tissue and sucking the cell sap, whereas the adult stage causes indirect damage by transmitting the Iris

yellow spot virus, which reduces both qualitative and quantitative loss [27]. This species are colonized in a wide variety of crops and weeds but predominantly preferred to onion [28, 29]. Our general result, a thrips population has appeared at the 9th SMW approximately fifty days of transplanting of onion seedlings and maximum colonization at the 15th SMW. This finding is consistent with Raut and coauthor [29], who reported that the thrips population appeared at the 9th SMW even though the seedlings were transplanted at different times, with two identical picks (12th and 15th SMW) observed in Jalandhar, Punjab. Crop season, host, geographical location, varieties, morphological and biochemical parameters, and climatic conditions all influence thrips population colonization. Furthermore, climatic factors play an important role in the development of thrips. Our findings indicated that thrips populations increased with maximum temperature and decreased with minimum temperature and relative humidity. The current findings are supported by Raut *et al.*, [29], who reported hot and humid climatic conditions favorable for linear growth. Parthenogenesis and sexual reproduction are the main factors for rapid development in a short time in this species [30]. Because of morphological differences in the crops, there was a significant difference in the development of the immature stage of thrips on cabbage, garlic, and onion cultivars [31]. *Lantana camara* is a well-known ornamental weed used to treat a variety of diseases and in various folk medicinal preparations [32, 33]. As major phytochemical functional groups, *L. camara* Linn contains essential oils, phenolic, flavonoids, carbohydrates, proteins, alkaloids, glycosides, iridoid glycosides, phenyl ethanoid, oligosaccharides, quinine, saponins, steroids, triterpenes, sesquiterpenoids, and tannin [34-37]. As a result, the presence of these phytochemicals demonstrates significant biological activities such as antipyretic, antimicrobial, antimutagenic, antibacterial, fungicidal, insecticidal, and nematocidal properties [38, 39]. These plants have insecticidal properties due to the presence of dodecanol, 1-eicosano, piperidine, and ethoxy [40], while lantanilic acids, camaric acid, and oleanolic secondary metabolites are responsible for nematocides

[41]. Furthermore, oleanonic acid, lantadene A, and oleanolic acid are toxic to aquatic arthropods but not to herbivorous arthropods [42]. Recently many researchers focused on identifying promising secondary metabolites in *L. camera* and studied its toxicity impact on various herbivores. Extract of *L. camera* had a significant impact on lepidoptera pest, sucking and stored grain pest [43-45]. Plant extracts are widely used in the pest management program in economic crops because of ecofriendly, did not develop resistance to a pest, have no phytotoxic effect, and are easily available in local areas. Therefore, we used *L. camera* in our study and it resulted in leaf extract 50 g L<sup>-1</sup> having a significantly higher percentage in decline thrips population than other those with higher concentration. Several botanicals also have been evaluated against the *T. tabaci* in onion growing areas. The *L. camera* leaf extract has potential to decline the *T. tabaci* population when extracted in cow urine [46, 47]. Bio-pesticides are compatible with synthetic pesticides which increase the efficacy against the pest in a sustainable manner. In the field, the application of *L. camera* leaf extracts with diamethoate. Our results show that the combination of botanical and synthetic insecticides i.e., *L. camera* leaf extract @ 50 g L<sup>-1</sup>, 100 g L<sup>-1</sup> and 150 g L<sup>-1</sup> with Dimethoate 30% EC @ 250 mL ha<sup>-1</sup> against the population of thrips and results revealed that the effective control thrips population as compared to the control (untreated). Dimethoate is a widely used organophosphate insecticide and acaricide. It was patented and introduced in the 1950s by American Cyanamid. Dimethoate is an acetylcholinesterase inhibitor that disables cholinesterase, an enzyme essential for central nervous system function. It acts both by contact and through ingestion. It is readily absorbed and distributed throughout plant tissues, and is degraded relatively rapidly [48]. Our results in agreements that application of diamethoate 30% EC @ 250 mL ha<sup>-1</sup> against thrips population found satisfactory results [49, 50].

### CONCLUSION

From the present investigation, it can be concluded that *Lantana camera* leaf extract

could be used in onion fields to manage *Thrips tabaci* sustainably. Thus, it offers more effective and diverse control options, as well as environmentally sound and cost-effective solutions.

**ACKNOWLEDGMENTS:** Authors are grateful to the Lovely Professional University, Phagwara, Punjab for providing all necessary facilities for conducting the research trials.

**CONFLICT OF INTEREST:** None

**FINANCIAL SUPPORT:** None

**ETHICS STATEMENT:** None

### REFERENCES

1. CSA. Agricultural Sample Survey report of the 2010/2011 (September-December 2010) Area and Production of Major Crops (Private Peasant Holdings, Meher Season). Central Statistical Agency, Addis Ababa, Ethiopia. 2011.
2. Kianian F, Marefati N, Boskabady M, Ghasemi SZ, Boskabady MH. Pharmacological Properties of *Allium cepa*, preclinical and clinical evidences; a review. Iranian J Pharma Res. 2021;20(2):107. doi:10.22037/ijpr.2020.112781.13946
3. Lee JT, Bae DW, Park SH, Shim CK, Kwak YS, Kim HK. Occurrence and biological control of post-harvest decay in onion caused by fungi. J Plant Pathol. 2001;17(3):141-8.
4. Wessal YHA, Hassan OK, Ammar MS Abdalla. Economic Evaluation of Some Botanical Oils for the Control of Onion Thrips (*Thrips tabaci* Lind.; Thysanoptera: Thripidae) and Onion Yield. Acta Sci Pharm Sci. 2019;3(8):124-8. doi:10.31080/ASPS.2019.03.0351
5. Soto-Rojas L, Rodríguez-Leyva E, Bautista-Martínez N, Ruíz-Galván I, García-Palacios D. Sequential and Binomial Sampling Plans to Estimate Thrips *tabaci* Population Density on Onion. Insects. 2021;12(4):331. doi:10.3390/insects12040331
6. Jones DR. Diseases of banana, abaca, and enslet. CABI, London, UK. 2000.
7. Bekele E, Azerefegne F, Abate T. Facilitating the implementation and adoption of IPM in Ethiopia. Proceedings of a Planning

- Workshop, ASAI/EARO, Nazerath, Ethiopia. 2006.
8. Loredó Varela RC, Fail J. Host Plant Association and distribution of the onion thrips, *Thrips tabaci* cryptic species complex. *Insects*. 2022;13(3):298. doi:10.3390/insects13030298
  9. Anonymous. Package of practices for cultivation of vegetables. Punjab Agricultural University, Ludhiana, Punjab (Ed. Thind SK). 2021:68-73.
  10. Nault BA, Hessney ML. Onion thrips control in onion, 2007. *Arthropod Manage Tests*. 2009;33:E20.
  11. Rani L, Thapa K, Kanojia N, Sharma N, Singh S, Grewal AS, et al. An extensive review on the consequences of chemical pesticides on human health and environment. *J Cleaner Prod*. 2021;283:124657. doi:10.1016/j.jclepro.2020.124657
  12. Shah R. Pesticides and Human Health. In: Nuro, A., editor. *Emerging Contaminants* [Internet]. London: IntechOpen; 2020. doi:10.5772/intechopen.93806. Available from: <https://www.intechopen.com/chapters/73921>
  13. Morse JG, Hoddle MS. Invasion biology of thrips. *Ann Rev Entomol*. 2006;51:67-89.
  14. Isman MB. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Ann Rev Entomol*. 2006;51:45-66.
  15. Dayan FE, Cantrell CL, Duke SO. Natural products in crop protection. *Bioorg Med Chem*. 2009;17(12):4022-34. doi:10.1016/j.bmc.2009.01.046
  16. Stoll G. *Natural Crop protection in the tropics: Letting information come to life*. Margraf Verlag; 2000. p.387.
  17. Talukder FA. Plant products as potential stored product insect management agents a mini review. *Emir J Agric Sci*. 2006;18:17-32.
  18. Dubey NK, Srivastava B, Kumar A. Current status of plant products as botanical pesticides in storage pest management. *J Biopestic*. 2008;1(2):182-6.
  19. Hazarika LK, Barua NC, Kalita S, Gogoi N. In search of green pesticides for tea pest management: *Phlogocanthus thyrsoflorus* experience. In: Ignacimuthu S, Jayraj S (eds) *Recent Trends in Insect Pest Management*. Elite Publication, New Delhi. 2008:79-90.
  20. Ogunleye RF, Ogunkoya M, OandAbulude FO. Effect of the seed oil of three botanicals, *Jatropha curcas* (L), *Helianthus annuus*(L.) and *Cocos nucifera*(L) on the maize weevil, *Sitophilus zeamais*(Mots.). *Plant Prod Res J*. 2010;14:14-8.
  21. Khan M, Srivastava SK, Syamasundar KV, Singh M, Naqui AA. Chemical composition of leaf and flower essential oils of *Lantana camara* from India. *Flavour Fragr J*. 2002;17(1):75-7.
  22. Khan M, Srivastava SK, Neetu J, Syamasundar KV, Yadav AK. Chemical composition of fruits and stem essential oils of *Lantana camara* from northern India. *Flavour Fragr J*. 2003;18(5):376-9.
  23. Ayalew AA. Insecticidal activity of *Lantana camara* extract oil on controlling maize grain weevils. *Toxicol Res Appl*. 2020;4:2397847320906491. doi:10.1177/2397847320906491
  24. Pung T, Srimongkolchai W. Toxic effects of *Lantana camara* crude extracts on *Spodoptera litura* (Fabr.). *Asian J Chem*. 2011;23(7):2863-5.
  25. Prema MS, Ganapathy N, Renukadevi P, Mohankumar S, Kennedy JS. Efficacy of different botanical extracts on *Thrips palmi* in cotton. *J Pharmacog Phytochem*. 2018;7(2):2824-9.
  26. Kumar D, Prakash D, Agrawal V, Nebapure S, Ranjan A, Jindal T. Bio-efficacy of Indian weed plants *Lantana camara* on cotton mealy bug (*Phenacoccus solenopsis*). *Plant Arch*. 2019;19(2):820-3.
  27. Pal S, Wahengbam J, Raut AM, Banu AN. Eco-biology and management of onion thrips (Thysanoptera: Thripidae). *J Entomol Res*. 2019;43(3):371-82. doi:10.5958/0974-4576.2019.00066.5
  28. Palomo LAT, Martínez NB, Johansen-Naime R, Napoles JR, Leon OS, Arroyo HS, et al. Population fluctuations of thrips (Thysanoptera) and their relationship to the phenology of vegetable crops in the central region of Mexico. *Fla Entomol*. 2015;430-8. doi:10.1653/024.098.0206
  29. Raut AM, Pal S, Wahengbam J, Banu AN. Population dynamics of onion thrips (*Thrips tabaci* Lindeman, Thysanoptera:

- Thripidae) and varietal response of onion cultivars against onion thrips. *J Entomol Res.* 2020;44(4):547-54. doi:10.5958/0974-4576.2020.00092.4
30. Fekrat L, Shishehbor P, Manzari S, Nejadian ES. Comparative development, reproduction and life table parameters of three populations of Thrips tabaci (Thysanoptera: Thripidae) on onion and tobacco. *J Entomol Soc Iran.* 2009;29(1):11-23.
31. Basri R, Ansari MS, Moraiet MA, Muslim M. Influence of host plants on biological parameters of Thrips tabaci. *Ann Plant Prot Sci.* 2019;27(1):31-6. doi:10.5958/0974-0163.2019.00006.5
32. Melanie M, Hermawan W, Kasmara H, Panatarani C. Physicochemical characterizations and insecticidal properties of Lantana camara leaf ethanolic extract with powder application. In: *AIP Conference Proceedings.* AIP Publishing LLC. 2020;2219(1):040002. doi:10.1063/5.0003200
33. Shah M, Alharby HF, Hakeem KR. Lantana camara: A comprehensive review on phytochemistry, ethnopharmacology and essential oil composition. *Lett Appl Nanosci.* 2020;9(3):1199-207. doi:10.33263/LIANBS93.11991207
34. Bhakta D, Ganjewala D. Effect of leaf positions on total phenolics, flavonoids and proantho-cyanidins content and antioxidant activities in Lantana camara (L.). *J Sci Res.* 2009;1(2):363-9.
35. Venkatachalam T, Kumar VK, Selvi PK, Maske AO, Kumar NS. Physicochemical and preliminary phytochemical studies on the Lantana camara (L.) fruits. *Int J Pharm Pharm Sci.* 2011;3(1):52-4.
36. Kensa VM. Studies on phytochemical screening and antibacterial activities of Lantana camara Linn. *Plant Sci Feed.* 2011;1(5):74-9.
37. Kalita S, Kumar G, Karthik L, Rao KV. Phytochemical composition and in vitro hemolytic activity of Lantana camara L. (Verbenaceae) leaves. *Pharmacologyonline.* 2011;1:59-67.
38. Verma RK, Verma SK. Phytochemical and Termicidal study of Lantana camara var. aculeata leaves. *Fitoterapia.* 2006;77(6):466-8.
39. El Baroty GS, Goda HM, Khalifa EA, Abd El Baky HH. Antimicrobial and antioxidant activities of leaves and flowers essential oils of Egyptian Lantana camara L. *Der Pharma Chem.* 2014;6:246-55.
40. Ayalew AA. Insecticidal activity of Lantana camara extract oil on controlling maize grain weevils. *Toxicol Res Appl.* 2020;4:2397847320906491. doi:10.1177/2397847320906491
41. Qamar F, Begum S, Raza SM, Wahab A, Siddiqui BS. Nematicidal natural products from the aerial parts of Lantana camara Linn. *Nat Prod Res.* 2005;19(6):609-13. doi:10.1080/14786410512331330594
42. Fatope MO, Salihu L, Asante SK, Takeda Y. Larvicidal activity of extracts and triterpenoids from Lantana camara. *Pharma Bio.* 2002;40(8):564-7. doi:10.1076/phbi.40.8.564.14654
43. Singh M, Naik JH, Pande H, Kaushal BR. Biological effects of a plant extract on gram pod borer, Helicoverpa armigera (Hübner)(Lepidoptera: Noctuidae). *J Env Bio Sci.* 2019;33(1):177-83.
44. Cathrine S, Lovejoy T, Rumbidzai CR. Effectiveness of tick berry (Lantana camara) in controlling larger grain borer (Prostephanus truncatus) in stored maize. *Asian Food Sci J.* 2022;21(3):10-8. doi:10.9734/AFSJ/2022/v21i330413
45. Feyisa B, Lencho A, Selvaraj T, Getaneh G. Evaluation of some botanicals and Trichoderma harzianum against root-knot nematode (Meloidogyne incognita (Kofoid and White) Chit wood) in tomato. *J Entomol Nematol.* 2016;8(2):11-8. doi:10.5897/JEN2015.0145
46. Singh DK, Verma TC, Aswal S, Aswani G. Effect of different botanical pesticides against Thrips tabaci on garlic crop. *Asian Agri Hist.* 2014;18(1):57-61.
47. Borhade GR, Sangeetha JS, Bhor GL. Efficacy of a novel phytopesticide against Thrips tabaci, an insect pest on onion. *Ann Plant Sci.* 2018;7(1):1949-51.
48. Dauterman WC, Viado GB, Casida JE, O'Brien RD. Insecticide residues, persistence of dimethoate and metabolites

- following foliar application to plants. J Agric Food Chem. 1960;8(2):115-9.
49. Patil LB, Patil CS. Bioefficacy of insecticides against onion thrips (*Thrips tabaci* Lindeman). J Pharmacogn Phytochem. 2018;7(1):958-61.
50. Singh H, Cheema HK, Singh R. Field evaluation of horticultural mineral oils and botanicals against bean thrips, *Megalurothrips distalis* (Karny)(Thysanoptera: Thripidae), in summer mung bean. Egypt J Biol Pest Control. 2020;30(1):1-8.